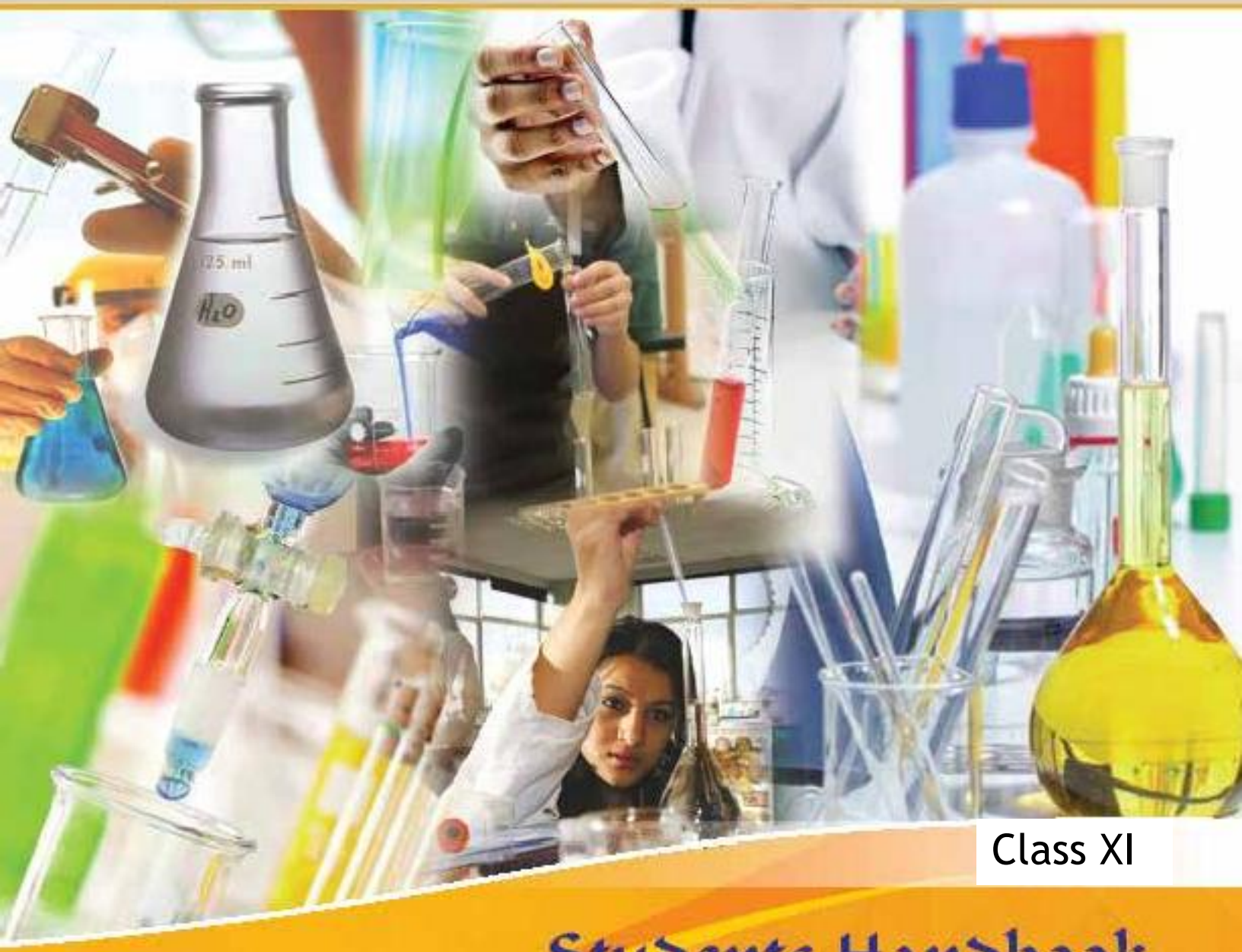
CBSE STUDY MATERIAL 2019

LABORATORY MEDICINE–I



Class XI

Students Handbook

Study Material

Central Board of Secondary Education
Shiksha Kendra, 2, Community Centre, Preet Vihar, Delhi-110301

Laboratory Medicine – II

Student Handbook, Class-XII

Price : `

First Edition :

Copies :

Paper Used : 80 GSM CBSE Water Mark White Maplitho

Published By : The Secretary, Central Board of Secondary Education, Shiksha Kendra, 2, Community Centre, Preet Vihar, Delhi-110092

Design & Layout : Public Printing (Delhi) Service
C-80, Okhla Industrial Area, Phase-I, New Delhi-110020

Printed By :

भारत का संविधान

उद्देशिका

हम, भारत के लोग, भारत को एक सम्पूर्ण [1]प्रभुत्व-संपन्न समाजवादी पंथनिरपेक्ष लोकतंत्रात्मक गणराज्य बनाने के लिए, तथा उसके समस्त नागरिकों को:

सामाजिक, आर्थिक और राजनैतिक न्याय,
विचार, अभिव्यक्ति, विश्वास, धर्म
और उपासना की स्वतंत्रता,
प्रतिष्ठा और अवसर की समता
प्राप्त कराने के लिए
तथा उन सब में व्यक्ति की गरिमा
[2]और राष्ट्र की एकता और अखंडता
सुनिश्चित करने वाली बंधुता बढ़ाने के लिए

दृढ़संकल्प होकर अपनी इस संविधान सभा में आज तारीख 26 नवम्बर, 1949 ई॰ को एतद्द्वारा इस संविधान को अंगीकृत, अधिनियमित और आत्मार्पित करते हैं।

---

1. संविधान (बयालीसवां संशोधन) अधिनियम, 1976 की धारा 2 द्वारा (3.1.1977) से "प्रभुत्व संपन्न लोकतंत्रात्मक गणराज्य" के स्थान पर प्रतिस्थापित।
2. संविधान (बयालीसवां संशोधन) अधिनियम, 1976 की धारा 2 द्वारा (3.1.1977) से "राष्ट्र की एकता" के स्थान पर प्रतिस्थापित।

भाग 4 क

मूल कर्त्तव्य

51 क. मूल कर्त्तव्य – भारत के प्रत्येक नागरिक का यह कर्त्तव्य होगा कि वह –

(क) संविधान का पालन करे और उसके आदर्शों, संस्थाओं, राष्ट्रध्वज और राष्ट्रगान का आदर करे;

(ख) स्वतंत्रता के लिए हमारे राष्ट्रीय आंदोलन को प्रेरित करने वाले उच्च आदर्शों को हृदय में संजोए रखे और उनका पालन करे;

(ग) भारत की प्रभुता, एकता और अखंडता की रक्षा करे और उसे अक्षुण्ण रखे;

(घ) देश की रक्षा करे और आह्वान किए जाने पर राष्ट्र की सेवा करे;

(ङ) भारत के सभी लोगों में समरसता और समान भ्रातृत्व की भावना का निर्माण करे जो धर्म, भाषा और प्रदेश या वर्ग पर आधारित सभी भेदभाव से परे हों, ऐसी प्रथाओं का त्याग करे जो स्त्रियों के सम्मान के विरुद्ध हैं;

(च) हमारी सामासिक संस्कृति की गौरवशाली परंपरा का महत्त्व समझे और उसका परिरक्षण करे;

(छ) प्राकृतिक पर्यावरण की जिसके अंतर्गत वन, झील, नदी, और वन्य जीव हैं, रक्षा करे और उसका संवर्धन करे तथा प्राणी मात्र के प्रति दयाभाव रखे;

(ज) वैज्ञानिक दृष्टिकोण, मानववाद और ज्ञानार्जन तथा सुधार की भावना का विकास करे;

(झ) सार्वजनिक संपत्ति को सुरक्षित रखे और हिंसा से दूर रहे;

(ञ) व्यक्तिगत और सामूहिक गतिविधियों के सभी क्षेत्रों में उत्कर्ष की ओर बढ़ने का सतत प्रयास करे जिससे राष्ट्र निरंतर बढ़ते हुए प्रयत्न और उपलब्धि की नई उंचाइयों को छू ले;

[1](ट) यदि माता-पिता या संरक्षक है, छह वर्ष से चौदह वर्ष तक की आयु वाले अपने, यथास्थिति, बालक या प्रतिपाल्य के लिये शिक्षा के अवसर प्रदान करे।

---

1. संविधान (छयासीवां संशोधन) अधिनियम, 2002 की धारा 4 द्वारा प्रतिस्थापित।

THE CONSTITUTION OF INDIA

PREAMBLE

WE, THE PEOPLE OF INDIA, having solemnly resolved to constitute India into a [1]**SOVEREIGN SOCIALIST SECULAR DEMOCRATIC REPUBLIC** and to secure to all its citizens :

JUSTICE, social, economic and political;

LIBERTY of thought, expression, belief, faith and worship;

EQUALITY of status and of opportunity; and to promote among them all

FRATERNITY assuring the dignity of the individual and the[2] unity and integrity of the Nation;

IN OUR CONSTITUENT ASSEMBLY this twenty-sixth day of November, 1949, do **HEREBY ADOPT, ENACT AND GIVE TO OURSELVES THIS CONSTITUTION.**

---

1. Subs, by the Constitution (Forty-Second Amendment) Act. 1976, sec. 2, for "Sovereign Democratic Republic" (w.e.f. 3.1.1977)
2. Subs, by the Constitution (Forty-Second Amendment) Act. 1976, sec. 2, for "unity of the Nation" (w.e.f. 3.1.1977)

THE CONSTITUTION OF INDIA

Chapter IV A

FUNDAMENTAL DUTIES

ARTICLE 51A

Fundamental Duties - It shall be the duty of every citizen of India-

(a) to abide by the Constitution and respect its ideals and institutions, the National Flag and the National Anthem;

(b) to cherish and follow the noble ideals which inspired our national struggle for freedom;

(c) to uphold and protect the sovereignty, unity and integrity of India;

(d) to defend the country and render national service when called upon to do so;

(e) to promote harmony and the spirit of common brotherhood amongst all the people of India transcending religious, linguistic and regional or sectional diversities; to renounce practices derogatory to the dignity of women;

(f) to value and preserve the rich heritage of our composite culture;

(g) to protect and improve the natural environment including forests, lakes, rivers, wild life and to have compassion for living creatures;

(h) to develop the scientific temper, humanism and the spirit of inquiry and reform;

(i) to safeguard public property and to abjure violence;

(j) to strive towards excellence in all spheres of individual and collective activity so that the nation constantly rises to higher levels of endeavour and achievement;

[1](k) who is a parent or guardian to provide opportunities for education to his/her child or, as the case may be, ward between age of 6 and 14 years.

---

1. Subs. by the Constitution (Eighty - Sixth Amendment) Act, 2002

Preface

I am very pleased to present the first edition of the text book in for the laboratory part of medical diagnostics.This is a new vocational course from Central board of Secondary Education.This is a part of the dream project of our Prime minister Mr.NarendraModiji

For skilling India. This is to ensure that all students who pass their 12th board examinations shall have the capability to get gainful employment because of their skill set.

This book has been written by Competent persons actively working in the various field of laboratory medicine –which includes histopathology,cytology,hematology,clinicalpathology,microbiology,blood banking etc. They are professors ,writers ,practising doctors and academicians.

This book shall have contents that shall cover the complete course curriculum for classes 11th to 12 th for the areas of Medical Diagnostics.Thewriteup of the book is fairly simple and shall help the student update his knowledge in the subject including all recent developments.He shall be able to self judge /asses his own competency through a set of questions given for self assessment.

I would like to thank CBSE vocational Unit ,who had been the driving force behind the development of this book. who has pains taxingly devoted so much of her time inensuring that it comes along in this fine form. My team of subject expert specially to mention from Safdarjung Hospitals .The current text has been prepared keeping in view the current requirements of the students and the latest updates in the relevant areas in a concise mannerusing simple language for increasing the comprehension.

Constructive and helpful suggestions from readers for the improvement of the book are welcome.

Chairman, CBSE

Acknowledgements

- ANITA KARWAL, IAS, Chairperson , CBSE

Editing & Coordination

- Dr. Biswajit Saha, Director (Skill Education & Training), CBSE

Content Developed by

- Dr Juhee Chandra -Lab Director at Mediaid Diagnostics
- Dr.Rashmi Vohra -Director Vohra Diagnostics
- Dr Geetika Khanna, Professor of Pathology, CIO Laboratory, VMMC & Safdarjung Hospital.
- Dr. Neeraj JainMBBS. MD.Director, Jain Diagnostics

Contents

I

UNIT - 1

INVESTIGATION URINE & FAECES ANALYSIS

OVERVIEW

This unit will provide the student information about the scope of and the organization of a clinical pathology laboratory. It will help to understand the relevant terms, procedures and working of equipments pertaining to urine and faeces investigations.

Organization of a Clinical pathology Laboratory:

The personnel needs of a laboratory depends on overall work load and the different types of materials to be processed. Assuming that the laboratory consists of routine sections, the employees would include a laboratory head, skilled employees in the form of technicians to supervise the different sections, and unskilled employees in the form of laboratory assistants. Related areas omitted in this example should have close communications with the other departments, but maintain separate and distinct supervision.

The Chief of the Laboratory should be a trained pathologist.

The technicians should have a diploma in medical laboratory technology from a recognized institution. They are responsible for specimen collection, preparation and test validations.

Support Staff include clerical and secretarial workers in the laboratory. Physical Infrastructure of the laboratory must be well designed and conveniently located to enable the professional and support personnel to perform their duties effectively. It must contain four definitely separated areas:

- Reception.
- Specimen collection room.
- Processing area.
- Reporting room.

KNOWLEDGE AND SKILL OUTCOMES

i. To understand the scope of clinical pathology.

ii. To know the organizational structure of a clinical pathology laboratory.

iii. To know the relevant terms, procedures and working of equipments pertaining to clinical pathology.

RESOURCE MATERIALS

i. a. Text Book of Medical Laboratory Technology, Praful B. Godkar, First edition.

ii. b. urine Analyzer insert.

iii. c. A Textbook of Biochemistry by Harbans Lal.

DURATION

LEARNING OUTCOMES

After completing this unit the students should be able to

1. Demonstrate knowledge, comprehension, and application of general techniques in the areas of:
   - Specimen accessioning.
   - Manual methods.
   - Routine general examination.
   - Microscopic examination.
2. Set-up, operate and maintain routine instruments.
3. Solve basic problems associated with reagents and methods of general techniques.
4. Apply principles of lab safety in completing all laboratory work.
5. Ensure quality control while performing general procedures.

INTRODUCTION TO urine examination

Urine is one of the most easily obtained specimens examined in the laboratory, and examination of the urine not only provides information about the functioning of the kidneys and possible abnormalities of the urinary tract, but may also lead to the diagnosis of various systemic diseases of the human body which are reflected by the presence of several abnormal substances in the urine.

PURPOSE OF THE EXAMINATION

To perform complete urine examination

LIS OF EQUIPMENT REQUIRED FOR SETTING UP OF A LABORATORY

1. Microscope
2. Centrifuge

3. Urine strips
4. Spirit lamp
5. Testtubes
6. Reagents for various manual tests slides/coverslips etc.

COLLECTION OF SPECIMEN

a) Early morning urine

The best urine specimen for routine analysis is collected in the morning. It is usually concentrated and has an acid pH. Casts and cells are poorly preserved in dilute or alkaline urine and traces of dissolved substances such as protein and sugar can be missed if the urine is very dilute.

b) Random urine

This specimen can also collected at any time and is convenient for the patient and is suitable for most screening purposes.

c) Preservative Used

For routine analysis, no preservative is required but the urine is best examined fresh. Bacterial growth will ruin a specimen if analysis delayed for more than 3 hours. Refrigeration is the best way to preserve it if analysis is delayed. Refrigeration for more than 24 hours is not recommended.

d) Container for urine collection

The container used must be thoroughly clean and free from any detergent or disinfectant residue since the oxidants contained in such cleaning agents may cause the test areas for glucose and blood to indicate false positive results. After the urine is collected, the container should preferable be sealed.

1. CLINICAL SIGNIFICANCE

Routine urine examination is performed mainly for two purposes

To find out metabolic or endocrine disorders in the body (e.g. Normal urine does not contain bilirubin or sugar. Presence of bilirubin in urine, indicates metabolic disturbance of bilirubin and sugar in urine is a indicator of diabetes mellitus i.e. deficiency of insulin, an endocrine disorder.

To detect intrinsic conditions that adversely affect kidney or urinary tract. Diseased kidneys cannot function normally in regulating the volume and composition of body fluids and also in maintaining acid-base balance and homeostasis.

Structural elements such as leukocytes, red blood cells and casts from the lower urinary tract may appear in urine. Substances normally retained by kidneys or excreted by kidneys in small mounts may also appear in large quantities and substances normally excreted maybe retained which is indicated by increased values in the blood e.g. Urea, Creatinine

3. The expected changes in the composition of urine stored at room temperature are as follows :-

a. Lysis of red blood cells by hypotonic urine
b. Decomposition of casts
c. Bacterial multiplication
D. Decrease in glucose level, due to bacterial growth
e. Formation of ammonia from urea by the action of bacteria (and the nature of urine changes to alkaline)

4. METHODS

The various aspects considered in the complete examination of urine are as follows :-

Physical examination of urine

Chemical examination of urine

Microscopic examination of urine

PHYS Following aspects are studies for the physical examination

Volume (Optional)

Colour

Appearance

Odour

Specific gravity

Reaction (pH)

VOLUME

A. For an adult, the normal average daily volume of urine is about 1200-1500 ml. More urine formation takes during the day than at night. However the normal range for 24 hours may be 600-2000 ml.

B. Polyuria is an abnormal increase in excretion of urine volume (>2500ml) as in the diabetes mellitus and diabetes insipidus.

C. Oliguria is a decrease in urine excretion (<500ml). The term anuria means the complete suppression of urine formation in spite of high fluid intake.

CLINICAL IMPORTANCE

A) POLYURIA :-Abnormal increase in urine volume > 2500ml/24 hours as in diabetes mellitus and diabetes insipidus.

B) OLIGURIA :-Decrease in urine volume <500ml/24 hours. Observed in renal and post-renal conditions.

C) ANURIA :- Complete suppression of urine formation as in renal failure.

COLOUR

COLOUR	FINDING (PATHOLOGIC / NON PATHOLOGIC)
Pale Yellow	Normal Finding
Yellow, Dark yellow, Brownish yellow to Orange	May be due to presence of water soluble (direct) bilirubin-Hepatic, post hepatic conditions/intake of the following : Food (yellow) colour, nitrofuration, Vitamin B-complex, senna, serotonin, pyridium and conc. urine
White	Presence of – a) Chyle (Chyluria) b) Pus (many WBCs)/Phosphates
Pink to Red	Presence of a) Haemoglobin (haemoglobbinuria) b) Myoglobin (Myoglobinuria) c) Acute Febrile disease d) Red Blood Cells (hematuria), Renal disease / Excretion of Red Urine after eating beets (Inherited metabolic sensitivity)
Brownish Black	Presence of – • Homogentisic acid (Alkaptonuria) A rare disorder. • Melanin (Malignant melanoma)/ Intake of the following :- Chloroquine, iron compounds, hyrdoquione, levodopa, metronidazole, qunine, resorcinol, nitrofurnation, etc.
Blue to green	Presence of – Biliverdin Pseudomonas infection/Intake of the following : Methylene blue, Azure-A acriflavine, phenylsalicylate

5.1.cAPPEARANCE

Normal urine colour varies from light yellow to deep amber. Urine colour sometimes may vary depending upon the diet and fluids, if any, consumed by the patient. The colour of urine is sometimes related to a pigment called “urochrome”. The degree of colour also depends on whether the specimen is concentrated of dilute.

Normal urine is usually clear. If the pH is alkaline, may be observed due to the precipitation of phosphates. Such urine should be centrifuged before analysis. Turbidity due to presence of chyle (chylomicrons) cannot be centrifuged, but required filtration using a special cellulose filter having <o.1 mm diameter and is confirmed with the help of ether test which dissolves the chylomicrons.

a. Normal urine is usually clear :it may appear cloudy if amphorous phosphates are present in alkaline urine or amorphous urates in acidic urine. Amorphous phosphates form white precipitate which dissolves when heated.

b. Urine may appear cloudy or turbid by the presence of leucocytes and epithelial cells.

c. This can be confirmed by microscopic examination. Bacteria can also cause cloudiness and mucous gives hazy appearance to urine. Fat and chyle give urine a milky colour. Presence of RBC may give urine turbid and smoky appearance.

5.1.dODOUR

Normal fresh urine has a mild odour of ammonia. Presence of ketone bodies gives urine a sweety or fruity smell. Contaminated urine with bacterial may give pungent shell due to the formation of ammonia. The urine of a infant with phenylketonuria gives musty odour.

TEST DONE ON DX URINE ANALYZER

a) pH
b) Specific Gravity
c) Glucose
d) Bilirubin
e) Urobiliinogen
f) Ketones
g) Nitrite
h) Blood Leucocytes

QDX urine test reagent strips for urine analysis and dip & read test strips are used as an in vitro diagnostics aid using urine specimen. The strips can provide qualitative & semi-quantitative determination.

PROCEDURE

1. Remove the set strip from the bottle after checking the expiry date and re-cap bottle immediately.
2. In case of discolouration or darkening of the reagent areas do not use the strip.
3. Dip the test strip completely for no more than I sec. in fresh well mixed uncentrifuged urine.
4. Remove the strip along the rim of the container to remove excess urine.
5. Blot one side of the test strip on absorbent paper on one side to remove remaining urine and prevent missing of chemicals.
6. Hold the strip up horizontally and compare the colours developed with the colour chart on the bottle label.
7. The reading time is 60-120 sec is critical for optimal results.

STORAGE

The strips must be kept in the original bottle only and should not be used beyond the expiry date, each strip can be used only once. Dessicant should not be removed from the body. The strip should be stored at temperature between 2O C-300 C.

5.1.eSPECIFIC GRAVITY

METHOD :- not there in material

PRINCIPAL :- Electrolyte (MX) in the form of salt in urine the reacts with Polymethylvinyl Ether and Maleic Acid (OOH) which are weak acid ionic exchanger. The reaction produces Hydrogenous ionogen which reacts with the pH indicator and produces a color change.

Result

Normal range in urine 1.020-1.030

Visual test range 1.00-1.030

Instrument test range 1.005-1.030

CLINICAL SIGNIFICANCE

A. Specific gravity at a constant temperature is the ratio of the weight of the volume of the urine to the weight of same volume of distilled water. The specific gravity determination of urine is used to measure the concentrating and diluting power of the kidneys. The specific gravity of urine varies throughout the day and the normal range for random urine specimen is 1.003 to 1.035. The range for a 24 hour urine specimen is 1.015 to 1.030.

B. Polyuria is observed in both diabetes mellitus and diabetes insipidus. But in diabetes insipidus, the specific gravity of urine is low while in diabetes mellitus the specific gravity is high due to presence of glucose in the urine.

C. HYPERSTHENURIA :- It is the condition in which orine has high specific gravity, High specific gravity of urine is observed in various conditions such as dehydration, eclampsia, proteinuria, diabetes mellitus and lipoid nephrosis.

D. ISOSTHENURIA :- Excretion of urine with fixed specific gravity of 1.010. It is the indicator of poor tubular reabsorption.

E. HYPOSTHENURIA :-Urine excretion with constantly low specific gravity (1.0007). It is found in pyelonephritis, hypertension, protein malnutrition and diabetes insipidus. Diuretic medicines, and natural diuretics such as alcohol and coffee also cause excretion of urine with low specific gravity.

5.1.fREACTION (pH)

MANUAL METHOD:

Use a narrow range pH paper, In some clinical situations, measurements of approximate pH within+0.5 pH units using a narrow-rang pH paper may be very helpful.

(a) PROCEDURE

Using pH paper

Put a drop of urine on a portion of pH indicator paper. The color obtained is compared with a standard chart. For checking the reliability of the pH paper cross check the pH of buffer solutions of known pH values having acidic and alkaline pH ranges.

Result :-Normal urine pH ranges from 4.5 to 8.0. The pH values are reported for example as 6.0 if pH paper is used shows as 6.1.

Interpretation and quality control

Urine pH is usually acidic in normal people, especially non vegetarians and is usually alkaline in vegetarians.

An early morning urine pH < 5.5 indicates that renal tubular acidification mechanism is intact. As a quality control measure, use certified reference buffers (commercial source), one in acidic rang, say, pH 4.0 and the other in alkaline range, preferably pH 9.2 to check the reliability of the pH paper used.

Always use a pH indicator paper before the date of expiry. Do not use outdated pH papers. Always close the bottle containing the pH paper tightly.

ANALYSER METHOD :-

PRINICIPLE :- This test is based on the double indicators (Methyl red and Bromothymol blue) which give colors ranging from red orange through green to blue covering the urine pH range of 5-9.

a) Results

Record the reading from the instrument display and enter the values in the specific register.

Urine pH : Normal Range : 6-7

QDX urine Strip's Measuring Range :

Visual Test range :- 5-8.5

Instrument Test Range :- 5-9

CLINICAL SIGNIFICANCE

A. The freshly voided normal urine is usually slightly acidic and its pH may range from 4.6-7.0. A high protein intake ingestion of acidic fruits produces acidic urine. Respiratory acidosis, metabolic acidosis (diabetes ketosis, starvation, severe diarrhea) produces acidic urine. Urinary tract infections caused by Escherichia Coli result in acidic urine.

In respiratory alkalosis (Hyperventilation) and in metabolic alkalosis (excessive vomiting) alkaline urine is excreted. A diet that is high in vegetables and citrus fruits causes an alkaline urine. Urinary infections caused by Proteus and Pseudomonas may cause alkaline urine.

5.2 CHEMICAL EXAMINATION OF URINE FOR ABNORMAL CONSTITUENTS

The routine chemical analysis for abnormal constituents of urine includes following investigations :

a) Glucose
b) Ketone bodies
c) Proteins
d) Blood
e) Bile salts
f) Bile pigments
g) Urobilinogen
h) Nitrite

5.2.aGLUCOSE

Manual Method :-

Sugar : Benedict's Test

a) **PRINCIPLE :-** Urinary sugars when boiled in Benedict's Reagent reduce copper sulphate to reddish cuprous oxide precipitate in hot alkaline medium, the intensity of which is proportional to the amount of sugar present in the urine. The results are reported as 1+, 2+, etc depending upon the colour and intensity of the cuprous oxide precipitate.

b) **Reagent :-** Dissolve 17.3 g of crystalline copper sulphate in about 800 ml of distilled water, then add 100 g of sodium carbonate, mix to dissolve and finally add 175 g of sodium citrate. Mix coloured bottle at 250-350C. Stable for one year.

c) **Procedure :-** To 5 ml of Benedict's reagent taken in an 18 x 150 mm glass tube, add 8 drops (0.5ml) of urine, mix well and boil for 2-3 minute over the flame. Cool the tube and observe for any colour change.

Determination of glucose in urine by using QDX Urine Analyzer

PRINCIPLE :- Glucose is oxidized by Glucose oxidase to form glucuronic acid and hydrogen peroxide. Hydrogen peroxide releases neo-ecotypes oxide (O) under the action of peroxidase (O) oxidizes potassium iodide which produces the color change.

Sensitivity 50-100mg/dl. Visual and instrument tests range : Negative – 100mg/dl

Reactive Ingredients : 1.7% w/w glucose oxidase (microbial.123 U); 0.2% w/w peroxidase (horseradish 203 U); 0.1% w/w potassium iodine; 71.8% w/w buffer, 26.2% w/w nonreactive ingredients.

INTERPRETATION OF THE RESULTS

No change in the original colour of Benedicts's solution – Negative

Solution appears pale green and slightly cloudy – Trace

Definity cloudy green 1+0.5%

Yellow to orange precipitate 2+-01%

Orange precipitate 3+-1.5%

Brick Red precipitate 4+ (≥2g/dl)

False positive reactions are known to occur due to presence of non-carbohydrate substances like ascorbic acid, homogentisic acid, creatinine and uric acid.

Reducing sugars like lactose, galactose, fructose and pentoses will also give a positive reaction. The dipstick technique is specific for glucose and eliminates the false positive reaction due to the substances mentioned above.

CLINICAL SIGNIFICANCE

The normal renal threshold for glucose is 180mg/dl. When the glucose exceeds the normal threshold the renal tubules cannot reabsorb all the filtered glucose and then glycosuria occurs. Glycosuria is seen in following conditions.

a. Diabetes Mellitus
b. Endocrine Disorder – Diabetes Mellitus, Cushing's syndrome, Pancreatic tumors, Hyperthyroidism, & Hyperpituitarism.
c. Phaeochromocytoma, Carcinoma of Pancreas, Pancreatitis.
d. Central Nervous System Disorders, Brain tumors, asphyxia, burns, infection, Myocardial infarction.
e. Liver disease, Glycogen storage disease, obesity.
f. Pregnancy : Reduced threshold for Glucose.
g. Aged : Glucose intolerance.
h. Glycosuria without huperglycemia-renal tubular dysfunction

OTHER Renal glycosuria, Alimentary glycosuria.

5.2.(b)KETONE BODIES

MUNUAL METHOD

Ketone bodies-Rothera's test

The three main ketone bodies are acetone, acetoacetic acid (diacetic acid) and betahydroxy butyricacid. Testing for ketone bodies should be done on fresh urine or the specimen kept at 4.c.

Principal

Acetone and acetoacetic acid react with sodium nitroprusside in the presence of alkali to produce a purple color.

Sensitivity: 10-20 mg/dl, Visual Test Range, Negative-300mg/dl, Instrument Test Range: Negative-150mg/dl.

Reactive ingredients: 5.7% w/w sodium nitroprusside: 29.9% w/w nonreactive ingredients. 64.4% w/w buffer.

Procedure

To 5.0 ml of urine taken in 18 x 150mm glass tube. Saturate it with solid ammonium sulphate. Add 0.5 ml of 1% sodium nitroprusside solution. Mix well. Add ammonia solution along the side of the tube so that it layers on top of the urine.

Result

If acetone and diacetic acid are present, then apurple (permanganate calomel red) colour will form at the junction of the two layers within 30-60 seconds. The result can be graded from trace to 3+ based on the intensity of the colour formed as detailed below.

No change in colour-Negative

Pinkish rang - +

Red ring - ++

Deep purple ring - +++

Interpretation add quality control

Ketone bodies are intermediary products of fat metabolism and their presence in the blood and then in the urine are indications that the metabolism is dis disordered or incomplete. This is associated with metabolic acidosis. This occurs in poorly controlled diabetes mellitus and also in starvation.

Normal urine does not contain methyl ketone. Weak false positive reactions may occur if the urine contains L-dopa and phenyl pyruvic acid.

If there is suspicion of a false positive test, heat the urine in a test tube in a Bunsen burner flame for one minute, allow to cool and repeat the Rothera's test. Heated urine.

5.2.(h)URINE FOR NITRITE using QDX Urine Analyzer

PRINCIPAL :- Nitrite in urine & aromatic amino sulphanilamide are diazotized to from diazonium compound. The diazonium compound reacts with tetra hydro benzo quinolin 3 phenol causing the color change.

Normal range in urine zero

Visual & Instrument test range – Negative

Sensitivity – 0.25-0.4 mg/dl

Abnormal value means

1. UTI
2. Bacterial infection

MICROSCOPY

PURPOSE OF THE EXAMINATION

To perform microscopic examination of urine.

CLINICAL SIGNIFICANCE

The microscopic examination is a valuable diagnostic tool for the detection and evaluation of renal and urinary tract disorders and other systemic diseases.

METHOD

PRINCIPAL

The microscopic elements present in urine are collected in the form of deposits by centrifugation. A small drop of the sediment is examined by making a cover slip preparation under microscope.

REQUIREMENTS

a. Test tubes
b. Slides
c. Cover slip
d. Pipettes/Droppers
e. Microscope
f. Centrifuge machine

SPECIMEN

Freshly collected midstream clean catch urine.

PROCEDURE

a. Shake the urine container well and take about 5 ml of urine in the centrifuge tube.
b. Centrifuge the tube for 5 minutes at 2500 rpm.
c. Discard the supernatant quickly and completely into another tube (clear supernatant can be used for proteins determination) Resuspend the deposit by shaking the tube.
d. Take a clean glassslide and mark it with patient identification number.
e. Please one drop of deposit on a slide: cover it with a cover slip.
f. Observe the slide under low power objective and then under high power objective lens of the microscope by lowering the condenser to minimize light.
g. Record the findings.

MICROSCOPIC FINDINGS

The various findings can be observed by microscopy on the sediment may be as follows.

LEUCOCYTES - can also be detected by analyser method

Normal pus cells in urine : 2:3/hpf

Abnormal finding: 5/hpf

2% acetic acid can be added to the slide to accentuate the nuclei of leucocytes.

BY ANALYSER METHOD

Normal range in urine 0-10 WBC's/ul

Measuring range 0-500 WBC's/UL

Sensitivity 5-15 Lenko/ul

EPITHELIAL Cells - Squamous, tubular, transitional epithelial cells

Normal ; male : 2-3/hpf: Femal : 2-5/hpf

Abnormal :>5/hpf

ERYTHROCYTES - Presence of RBC's with intact membrane or dysmorphic RBC with half moon shape or irregular shape with crenated margins. Yeasts cells can be mistaken for RBC's Yeast

Normal : 1-2/hpf

Normal : Absent

The various abnormal casts found in urine specimen are as follows.

a. Granular casts
b. Hyaline casts
c. Red cell casts
d. White cell casts
e. Epithelial cell casts
f. Waxy casts
g. Fatty acids casts

CRYSTALS

CRYSTALS FOUND IN ACIDIC URINE

a. Uric acid crystals: are rosette shaped Can be present normally. Also seen in gout, chronic nephritis
b. Calcium oxalate crystals: are envelope shaped Can be present after ingestion of tomatoes, spinach, oranges, also seen in diabetes milletus, liver diseases
c. Amorphous urates: have no clinical significance
d. Sodium urates: have no clinical significance

e. Calcium sulphate : are thin colorless needles – have no clinical significance
f. Hippuric acid : are elongated prisms or plates-have no clinical significance
g. Cystine : are colorless hexagonal plates with equal or unepual sides-seen in cystinosis
h. Tyrosine : are fine refractile needles, occurring in clusters or sheaves-seen in tyrosinosis
i. Leucine : are oily, refractile spheroids: Seen in severe haepatitis, maple syrup disease.
j. Cholesterol : are transparent plates with notched corners seen in nephritis, nephritic condition, chyluria.

CRYSTALS FOUNDI N ALKALINE URINE

a. Tripe phosphate (ammonium magnesium phosphate) : are colorless prisms with 3-6 sides seen in normal urine, chronic cystitis, pyelitis
b. Amorphous phosphate : are present in granular form – have no clinical significance
c. Calcium carbonate : are colorless in the form spherical, dumbbell shape or granular form have no clinical significance
d. Ammonium biurates : are yellow brown spherical bodies with or without irregular spindles

MUCUS THREADS

Normally-absent

Abnormally : in UTI

OVAL FAT BODIES AND FAT DROPLETS

Normally-absent

Abnormally in nephritic syndrome, diabetes milletus, chronic glomerulonephritis, fat embolism

SPERMATOZOA

Normally – after coitus

BACTERIA:

Normally-absent

Abnormally-UTI

YEAST CELLS-yeast cells are ovoid and often with buds

Abnormally : UTI (ESPECIALLY IN DAIBETICS)

i) PARASITES Normally absent. The parasites which can be seen are

j) a. Trichomonas vaginalis trophozoites
k) b. Enterobius vermicularis ova
l) c. Schistosoma haematobium ova
m)
n) ARTIFACTS THAT CAN BE SEEN IN URINE:
o) 1. STRACH CRYSTALS
p) 2. FIBRES
q) 3. OIL DROPLETS
r) 4. HAIR
s) 5. AIR BUBBLES
t) 6. TALCUM POWDER PARTICLES
u)
v) 6. POST EXAMINATION SAMPLE STORAGE

Urine specimens are stored at room temperature for 24 hrs. after which they are discarded following rule of biohazardous waste management.

STANDARD OPERATING PROCEDURE ON STOOL ROUNTINE EXAMINATION

1. PURPOSE OF THE EXAMINATION

To perform routine stool examination

2. CLINICAL SIGNIFICANCE

Most of the parasites and bacterial pathogens causing gastrointestinal infections primarily involve the intestine. The laboratory diagnosis if gastrointestinal infections is mainly based on examination of stood specimens.

3. METHODS

The various aspect involved in the stood examination are as follows

Gross or physical examination – color, consistency, Blood, Mucus and Adult Parasites or body parts of parasites.

Gross or physical examination – color, consistency, Blood, Mucus and Adult Parasites or body parts of parasites.

Chemical examination – Reaction (Ph) occult blood Mucus and Adult Parasites or body parts of parasites.

Microscopy –Trophozoites and cysts of protozoa's Larvae and Ova of Nematodes and Cestodes, Plant cells, meat fibers, Crystals, Fat globules, Pus cells Erythrocytes, bacteria and yeast cells.

4. SPECIMEN

Stool specimen is collected for the diagnosis of GIT infections/other.

Gastrointestinal diseases e.g. Steatorrhoea.

Following precautions must be taken before collection stool specimen for routine examination & culture.

a. Stool should be collected prior to antibiotics : barium meal or mineral oil is given to the patient.

b. Do not contaminate faeces with urine.

c. Collect at least 3 specimens on 3 consecutive days.

e. in case of delay in analysis mix the stood specimen with transport media like

(Amies transport medium or buffered glycerol saline), while for parasitic Examination stool can be collected in 10% formal saline (3 parts formal saline and one part stood) or PVA (poly vinyl Alcohol).

f. Formal saline preserves helminthic eggs & larvae, while PVA is an excellent preservative for protozoan trophozoite stage.

g. before collection stood for occult blood test, the patient is asked to avoid for 3 days, the following. Red meat (Beef, lamb & liver) Vitamin C excess of 250mg/ day, citrus fruits & juices. High peroxidase containing fruits & vegetables including turnips, radish, Horseradishes, broccoli, & Cauliflower.

5. PROCEDURE

GROSS OR PHYSICAL EXAMINATION : Observe the stood for

Color : Light to dark brown, bright red, Black, Clay colored, Fresh blood, white colored

Consistency : Well formed, Solid, Semisolid, Liquid, Rice water stools, Pale, Bulky, Frothy and mucoid.

Blood

Mucus

Adult Parasites or body parts of parasites

CHEMICAL EXAMINATION OF STOOD

A. REACTION (pH)

CLINICAL SIGNIFICANCE

Normally stool is slightly acidic, Neutral or Slightly alkaline. (Range-5.8 to 7.5) Check the pH of stood using pH paper strip and note down the findings.

METHOD

pH paper strip method

6. PRINCIPAL

Indicators used in the test area are methyl red (pH range 4.4 to 6.2 color change from red to yellow) and bromothymol blue (pH range 8.0 to 9.6, color change from yellow to blue). When test strip is touched to the stood sample, color of the strip changes according to the pH of the stood.

REQUIREMENTS

a. pH paper srips (ranging from pH 2.0-10.5)

PROCEDURE

a. Dip the test strip in stool sample

b. Remove the strip from the sample and observe the color of the test strip (changes form orange to yellow and green through blue depending on the pH of the stood)

c. Compare the color with the corresponding color chart on bottle

d. Record the findings in the specific register.

RESULTS AND INTERPRETATION

Yellow color (no change in the color of the strip) Acidic Green through blue-Alkaline

B. STOOL OCCULT BLOOD

Theperoxidase activity of haemoglobin decomposes H2 O2 and liberates actives O2 which oxidizes the organic compound benizidine to give blue color.

HEMOSPOT test cards

Dropper bottle containing developer solution

Sample applicators

Positive control

STORAGE/STABILITY

Store the reagent at 20-30Oc, Cool please away from direct sunlight, fluorescent light U.V rays and moisture. Not refrigerate.

The reagents and test cards are stable till the expiry date mentioned on the label.

REAGENT PREPARATION

HEMOSPOT testcards consisting of a filter paper impregnated with the guaiac resin (Reactive surface), the Developer solution and the positive control and ready to use.

7. REQUIREMENTS

1. Mixing stix
2. Surgical hand gloves
3. Face mask

8. PROCEDURE

1. Piece the nozzle of the developer solution with a rust fee sharp pin or needle Retrieve the required number of test Label the cards with correct patient identity. Open the sample application window labeled A and B respectively, to expose the reactive surfaces of the test card.
2. By using the sample applicator provided in the kit spread a very thin layer of stool on the reactive surfaces on the window A similarly on window B from a different part of the stool.
3. Wait until the smeared sample has dried completely.
4. Turn over the test card.
5. Open the result window add one drop of developer to fields RA and RB (the reverse side of the sample smeared on the sample application windows respectively).
6. Observe for color change exactly at two minutes.
7. Even if one of the field's has a blue color, the test is positive for occult blood.

9. RESULTS AND INTERPRETATION

1. Negative For Occult Blood. No blue color indicates absence of occult blood in the stool.
2. Positive for occult blood.
   a. Trace blue colouration indicates presence of approximately 5mg/dl of occult blood in the stool.
   b. Strong blue colouration indicates significantly more than 5 mg/dl of occult blood in the stool.

Results are entered in specific registers, against specific samples, interpreted and released accordingly.

C. REDUCING SUBSTANCES IN STOOL BY BENEDICT'S TEST

1. CLINICAL SIGNIFICANCE

Causes of reducing substances in stool are as follows

Lactose intolerance (lactase deficiency in rotavirus infection of upper small intestine, leads to passage of lactose in stool)

2. PRINCIPAL

When benedicts reagent is heated with stool specimen the glucose present in stool reduces cupric to cuprous ions in Benedict's reagent. Alkaline medium provided in the reaction by sodium carbonate present in the reagent. The original blue color of Benedicts reagent changes to green, yellow, orange or brick red color according to the concentration of glucose present in stool.

3. REAGENT

Benedict's reagent

Composition

a. Copper sulphate

b. Na_2CO_3 (anhydrous)

4. REQUIREMENTS

a. Test tube

b. Test tube holder

c. Pipette

d. Bunsen burner

e. Centriguge

f. Dropper

g. Surgical hand gloves

5. PROCEDURE

a. Add 1 ml of stool to 2 ml of normal saline and mix thoroughly.

b. Pipette 5 ml of Benedict's reagent in a clean test tube and heart it in a Bunsen burner flame.

c. Then add sample 7-8 drops of saline suspension of stool sample to the tube Boil for 3-5 minutes and then cool the test tube.

d. Observe the color of the mixture and interpret the result.

6. RESULTS

Color change Reducing substances

1. No change in color (Blue) Absent
2. Color change to brick red Present

7. MICROSCOPIC EXAMINATION OF STOOL

APPLICATION

Sline & iodine wet mount for ova & cyst is simple & rapid method for diagnosis of gastrointestinal infection caused by either protozoa or helminthes.

PRINCIPAL

Saline preparation of stool specimen helps to demonstrate live protozoal & helminthic forms. While in iodine preparation the cysts of protozoa & helminthic eggs are stained brown & can by detected easily.

REAGENTS

a. Normal Saline (0.9%)

b. Lugol's iodine – (source-Himedia Laboratiories)

REQUIREMENTS

a. Slides

b. Cover slip

c. Droppers

d. microscope

e. Mixing sticks

PROCEDURE

a. Take a clean grit free slide.

b. Put one drop of normal saline oat one side of the slide & a drop of Lugol's iodine on another side of the slide.

c. Add on one drop of liquid stool to each of the normal saline & iodine drop. In case of solid or semisolid specimen mix small portion of stool to each of the drop.(Use separate applicator for mixing stool specimen in saline & iodine drop).

d. Cover the preparation with clean, grit free cover slip. Separate cover slips are used for saline & iodine preparation.

e. Observe both the preparation for presence of ova or cyst or live parasites under low & high power objectives of the microscope.

Figure 5.1 Student Microscope (Courtesy Thermoshandon)

Safety in the laboratory

- Treat all biological materials used in the laboratory as potentially infectious and pathogenic to humans.
- Laboratory coats must be worn by laboratory personnel at all times.
- All open cuts on hands and other exposed skin surfaces must be covered by gloves.
- Long hair should be tied back neatly, away from the shoulders.

- The lab should be well-ventilated and should strictly follow the regulations governing the acceptable limits. If solvents are used during practical sessions, the exhaust fan must be switched on.
- Proper disposal of hazardous wastes is a must. Disinfection by using 1% hypo and keeing it for a minimum of 20 min before discarding
- Every instrument used in the laboratory should meet electrical safety specifications and have written instructions regarding its use.
- It is advisable that flammable materials are stored with utmost care in appropriate storage cabinets that are designed for this purpose.
- Fire safety procedures are to be strictly adhered to. Safety equipment including first aid kits, fire extinguishers, fire blankets and fire alarms should be within easy access. A shower and eyewash should be.

1.10 Self-Assessment

1. Enumerate the steps in specimen receiving and accessioning.
2. Enlist the list of equipment required for setting up of a clinical pathology laboratory.
3. Outline the steps in routine processing of urine specimens.
4. What is automated processing?
5. What the various aspects of urine examination?
6. Why is volume of urine important?
7. What do the different colour of urine signify?
8. What chemical testing is done for urine when done manually?
9. How will you do a quality control for urine examination.
10. What is the method of disposal or urine samples.
11. How are stool samples transported to the laboratory?
12. What precautions are taken for stool occult blood sample collection?
13. How will you identify the various crystals and casts found in urine?
14. How will you identify the various ova / cyst found in stool Microscopy?

Sputum definition

It is a secretion that is produced in the lungs and the bronchi (tubes that carry the air to the lung), also is known as phlegm 2. This mucus-like secretion may become infected, bloodstained, or contain abnormal cells that may lead to a diagnosis S

Tracheobronchial sections are an inconstant mixture of plasma, water, electrolytes and mucin 4. As these mixture pas though the lower and upper respiratory tract, they become contaminated with cellular exfoliations, nasal and salivary gland secretions and normal bacterial flora of the oral cavity

Sputum collection

Drinking a lot of water and other fluids the night before the test may help to get the sample 2. To be asked to cough deeply and spit any sputum in a sterile cup 3. The sputum is ten taken to the laboratory 4. There, it is placed in a special substance (medium) under conditions that allow the organisms to grow

Sputum smear findings

This slide shows typical buccal squamous epithelial cells which are much larger than polymorphonuclear leukocytes (PMN) and take up most of the field in a high power view ●This cell is covered with chains of Gram positive cocci typical of normal oral flora such as peptostreptococci. (oil immersion, 1000x)

Physical properties of sputum

1 Appearance ●It may be described as liquid (serous), mucoil, purulent, bloody or combination of theses

2 Color ●Its color is determined by the material contained, and often color can indicate the pathological process ●Yellow color indicates pus and epithelial cells are present

3 Odor ●Usually no odor is present in normal and pathological sputum, but if bacterial decomposition has been taken place within the body or after expectoration, a variety of odor will be present

Miscellaneous findings

1 Cheesy Masses ●These are fragments of necrotic pulmonary tissue seen in such disease as pulmonary gangrene or tuberculosis

2 Bronchial Casts ●These are branching tree like casts of bronchi whose size varies with that of bronchi in which they are formed ●They are composed of fibrin and are white or gray color

3 Broncholiths (Lung Stones) ● They are formed by calcification of necrotic or infected tissues ● Chronic tuberculosis is the most common casue for their formation.

4 Dietrich's Plugs ● They are frequently observed in putrid bronchitis and bronchiectasis ● They are composed of cellular debris, fatty acids, crystals, fat globules and bacteria

HOW TO COLLECT A SAMPLE OF YOUR SPUTUM

If you are very sick, you may already be in the hospital. If so, the bedside nurse will help you cough up sputum to send to the laboratory for the test. If you have trouble coughing up sputum on your own, the nurse may have you breathe steam.

If you are sick at home, you will need to collect the sputum sample yourself.

Keep in mind that sputum from deep inside your lungs isn't the same as saliva. Sputum is mucus, and is usually colored and thick in consistency, especially when there is an infection in the lungs. Saliva comes from your mouth and is clear-colored and thin.

Plan to collect sputum first thing in the morning. This makes the test more accurate. Do not eat or drink anything in the morning before collecting your sput The sample can be refrigerated for up to 24 hours if needed. Do not freeze it or store it at room temperature.

Abnormal results mean that bacteria and white blood cells were seen in the sputum sample. The bacteria found will be either Gram-positive, or Gram-negative.

Common Gram-positive bacteria found by the test include:

- Staphylococcus
- Streptococcus
- Bacillus
- Listeria
- Enterococcus
- Clostridium

Common Gram-negative bacteria found by the test include:

- Cyanobacteria
- Spirochaetes
- Green sulfur bacteria
- Certain types of Proteobacteria

A normal test result means that there were very few white blood cells and no bacteria seen in the sputum sample.

Sputum stain for mycobacteria is a laboratory test performed on a sample of the patient's sputum (phlegm). It is also known as an acid-fast bacillus stain (AFB) or a tuberculosis (TB) smear. The test is commonly ordered by a doctor to find out if a patient has tuberculosis (TB) or another type of mycobacterial infection. ZN stain is done. This has been given in microbiology.

If your test results are normal (negative), this means no mycobacterial organisms were found.

If the test is abnormal, it means the stain is positive for one of the following organisms :

- Mycobacterium tuberculosis
- Mycobacterium leprae
- Nontuberculous bacteria
- Other acid-fast bacteria

5. Foreign Bodies ● In childer, they can be any small object a child may place it into his mouth ● In adults, they are either food particles or gastric contents aspirated during convulsion, during intoxication or operative anesthesia

6 Parasites ●They are extremely rare, but ascaris lumbricoides may be seen rarely.

Composition of sputum

Sputum Chemical Composition

Sold 5%, Water 95%

Solids are

DNA, Enzymes, a-antitrypsin, LDH, lysozyme, lactoferrin Lipids Proteins, Carbohydrates

UNIT - 2

Body Fluids

Overview

In this unit, we discuss various body fluids present in human body, their analysis and changes observed in these fluids in different diseases.

Various body fluids presents are :

Edema
Cerebrospinal Fluid
Pleural fluid
Peritoneal fluids
Pericardial fluids
Semen
Synovial fluid

Knowledge and skill outcomes

a) Anatomical site of fluids and physiological properties
b) Analysis of fluids
c) Variations in different diseases

Unit 2-Body Fluids	Outcomes
2.1 Edema	
2.2 Cerebrospinal Fluid	Understand Anatomical site, Physiological properties, pathological changes and estimation of cell counts (where relevant)
2.3 Pleural fluid	
2.4 Peritoneal fluid	
2.5 Pericardial fluid	
2.6 Semen	Method of collection, physical properties counts, morphology, biochemical estimation and interpretation
2.7 Synovial Fluid	Understand Anatomical site, Physiological properties, pathological changes and estimation of cell counts (where relevant)

EDEMA

It means swelling. It is defined as abnormal and excessive accumulation of fluid in the interstitial tissue spaces and serous cavities.

Following six mechanisms are responsible for edema.

1. Decreased plasma oncotic pressure
2. Increased capillary hydrostatic pressure
3. Lymphatic obstruction
4. Tissue factors (increased oncotic pressure of interstitial fluid, and decreased tissue tension).
5. Increased capillary permeability
6. Sodium and water retention.

List difference between Transudate and Exudate

Transudate	Exudate
1) Filtrate of blood plasma No change in capillary Permeability	Inflamed tissue oedema Increased capillary Permeability
2) Few cells	Many cells
3) Fluid Protein/Serum Protein<0.5	>0.5
4) Fluid LDH/Serum LDH<0.6	>0.6
5) Specific Gravity-Low	High
6) Examples- Congestive Heart Failure	Examples- TB Malignant effusion

CERBROSPINAL FLUID [C.S.F]

Three membranes or meninges cover the brain and spinal cord These are from out side to inside dura mater, arachnoid mater and pia mater which lies directly in contact with the brain. Most of the CSF is formed by the Choroid Plexus and circulates around the brain and spinal cord. It has a turnover rate of 20ml/hr.

Function of CSF

(A) Physical examination

A Appearance: Normal Cerebro Spinal Fluid. The specific gravity is 1.003-1.008. Normally the cells present are all lymphocytes, and their number is less than 5 per Cubicmm.

CSF is colour less, clear and any colour is abnormal. The most common cause of an abnormal colour is the presence of blood. This may come from trauma occurring during lumbar puncture.

In this case the first few drops will be the most heavily contaminated, and if the first 1 or 2 ml are collected separately the fluid collected after this may be almost clear. If there has been subarachnoid haemorrhage into the C.S.F. there will be blood throughout. If the fluid is bloody, centrifuge the specimen after taking a well mixed sample for cytologic study to see colour of fluid itself. A yellow colour in the C.S.F. is called Xanthochromia and may be due to haemorrhage some time before, the red blood cells in the C.S.F. having haemolysed and the hemoglobin liberated and slowly converted into bilirubin. The fluid is also often yellow when the spinal canal is blocked by a tumor, perhaps in part due to a great increase in protein level.

Turbidity: Turbidity is seen when there is a great increase in the number of cells i.e. to 400-500 polymorphs per cu.mm or more or when large numbers of organisms are present, e.g. in pneumococcal meningitis. Small numbers of R.B.C. cause smoky or opalescent appearance. Fibrin clots may form on standing in pathological fluids containing fibrinogen, which is usually only found when the protein is greatly increased. C.S.F. from patients with spinal tumour sometimes sets solid on standing. In tuberculous meningitis if the fluid is allowed to stand overnight a delicate clot like a cobweb often forms. This may take up the tubercle bacilli, which are more easily seen if the web is stained and examined microscopically. Hence it is useful, if possible, to leave part of the specimen to stand overnight while examining the rest immediately.

B. Cytological Examination:

Normal CSF contains very few cells; usually only 0-5 white blood cells per cu.mm and all those being small mononuclear cells – lymphocytes. Because there are so few cells, the fluid is often examined in the counting chamber undiluted. If the CSF appears cloudy one can make 1:20 dilution. 0.05 ml of CSF is added to 9.95ml of CSF diluting fluid. (2 percent v/v ascetic acid with 5 drops of 3gm/dl methylene blue. New bauer counting chamber is charged and left for about 5 min. to let the cells settle down.

Calculations

Leukocytes in CSF / per cumm (µ1)

$$= \frac{\text{Cells counted}}{0.9}$$

If CSF is diluted (1:20) then the calculation is

Leukocytes in CSF / per cumm (µ1)

$$= \frac{\text{Cells counted} \times 20}{0.9}$$

Differential Count:

1. Leishman stained smear: Here we make a smear of the centrifuged, deposit – and after it has dried, we stain it with Leishman stain or preferably a dilute solution of methylene blue. Cells in C.S.F. are commonly lymphocytes and neutrophils

Normal Cell Count In C.S.F: Adults 0-5 cells/cu.mm

Neonates 0-30 cells/cu.mm

Critical Value:>30 cells / cu.mm for any age.

Exercise

1. Describe the physical properties of normal cereberospinal fluid?
2. List the various causes of change in colour of cereberospinal fluid?
3. Match the following

(a)	Xanthochromiat	(1)	Traumatic collection of CSF
(b)	Blood in CSF	(2)	Tuberculosis
(c)	Cobweb formalities	(3)	Yellow colour of CSF

& 2.4 PLEURAL AND PERITONEAL FLUIDS

These fluids are found around lungs (Pleural) and in abdominal and pelvic cavities (Peritoneal)

The processing and reporting of the pleural and peritoneal fluids is essentially the same as that of CSF. The total cell count is done on Neubauer chamber using similar dilutions and the differential count is done on stained smears using same procedure.

The stains used may be Giemsa / leishman's / pap / H&E

FLUID CYTOLOGY :

The pleura and peritoneum is lined by mesothelium, hence mesothelial cells are normally found in the fluid.

Lymphocytic predominance is commonly seen in Tuberculosis.

Polymorphic (neutrophilic) predominance is suggestive of Bacterial infections.

Exercise

1. Give one reason for increased lymphocytes and neutrophils respectively in pleural fluid.

Pericardial Fluid

This is the fluid around the heart. Normally it is clear and straw coloured and about 20-50 ml in volume. It is analysis is similar to other body fluids. Pericardial fluid may be increased

in volume pathologically in case of congestive heart failure. Bloody pericardial fluid may be seen in traumatic tap, tuberculosis, bacterial pericarditis ets.

Exercise

1. List causes of bloody pericardial fluid.

SEMEN ANALYSIS

Semen is a fluid formed by testes and accessory male reproductive organs. It is composed of spermatozoa suspended in seminal plasma which provides a suitable nutrient medium and activates the spermatozoa to greater motility.

Method of collection: Specimens should be collected after a 3 day period of abstinence. Smoking and alcohol intake should be avoided during this period. Specimens should be collected in the laboratory itself. A clean and dry wide mouth bottle (50ml) is the recommended container for the collection of sample. The semen is collected by the patient and is delivered as soon as possible after collection, preferably within thirty minutes. The specimen liquefies usually within 15-30 minutes. It should be examined as soon as possible after liquefaction has taken place.

GROSS EXAMINATION :

Physical Characteristics :

Freshly ejaculated semen is a highly viscid, opaque, white or gray white coagulum with distinct musty or acrid odor. The coagulum will spontaneously liquefy within 10 to 20 mins to form a translucent, turbid, viscous fluid which is mildly alkaline with a pH of about 7.7. The specimen of normal viscosity can be poured drop by drop.

Coagulation and Liquefaction: Liquefaction should be complete within '30'mins.

Volume: Normal semen volume averages 2 to 6 ml

Chemical Examination:

Fructose Examination:

Fructose Test: (Qualitative):

Selewinoff's test:

Selwinoff's reagent:

Resorcinol - 50 mgs

D.H_2O - 70 cc

Conc. HCl - 30 ml

PROCEDURE:

1. To '2' ml of Selwinoff's reagent add 0.3 ml of semen and heat in boiling water bath for '5' min. A deep reddish color develops if fructose is present. And reported as positive / negative fructose test.

QUALITY CONTROL:

To be cross checked by a Sr. Technician / or duplicate test.

REFERENCE RANGE:

Normal study (WHO guidelines 1992)

Sperm count : 20-150 million / mlc

Sperm motility : >50% forward progressive motility (grade a+b)

Or

>25% rapid progressive motility (grade a)

Sperm morphology >30% Normal morphology

Volume : 2 to 6 ml

Viscosity : Normal / Absent

Ph : 7.2 -8.0

Sperm vitality / viability : > 75 % alive

WBC (absolute count) : < 1 million / ml

INTERPRETATION :

Normozoospermia : Normal study

Oligozoospermia : sperm count < 20 millions / ml

Asthenozoospermia : sperm motility < grade a or a+b

Oligoasthenospermia : count and motility less than normal

Azoospermia : absent sperms in the sample.

Microscopic Examination:

Sperm Count:

Following liquefaction of the semen, the spermatozoa can be counted in a hemocytometer chamber following dilution with diluting fluid in 1:20 dilution. (0.38 ml of diluting fluid and 0.02ml of the sample). After charging the chamber '2'min are allowed for immobilized sperms to settle. The spermatozoa in '4'sq.mm are counted under low power magnification.

Diluting fluid:

Sodium bi carbonate -5 gm

Formalin (neutral) -1 ml

Distilled water -99 ml

Sperms counted in 4 Sq. X 10 X 20 X 1000

Calculation = ----------

4

E) Motility:

To evaluate motility, a small drop of liquefied semen is placed on a microscopic slide and then covered with a cover slip and examined under the microscope with low and high power magnification. Motility can be evaluated by scanning several fields with a high dry objective, until a total of at least "200" spermatozoa have been observed. It is essential to focus entire depth of a given field so as to include non-motile sperm that have settled to the bottom of the medium. The sperm motility is graded as

a) Rapid Progressive motility: This is the motility along a linear track, covering a distance of at least20 micro mt./ sec (half the length of the spermatozoa).

b) Slow or sluggish progressive motility: This is the non linear motility

c) Non- progressive motility: spermatozoa move their tails but do not move forward.

d) Immotility (non- motile).

F) Sperm Morphology:Is evaluated by performing differential counts of morphologically normal and abnormal spermatozoa types on stained smears. Smears are prepared on clean, dry slides like blood films and the films are air dried and stained with Giemsa / Leishman stain. '200' spermatozoa should be examined under oil immersion and percentage of abnormal forms are recorded. Some abnormalities which may be seen in sperm are marked. Increase in head, size, tail, short tail, mid piece defects etc.

In addition to sperm morphology, the presence of RBC's, leucocytes and epithelial cells should be noted. Immature cells of germ line must be differentiated from macrophages and WBC's.

Exercise

1. What are the precautions to be taken while collecting semen sample?
2. Describe the physical characteristics of a normal semen sample.
3. Describe the procedure for evaluating motility of semen sample.

4. Describe the fructose test in semen.

5. What is azoospermia?

SYNOVIAL FLUID

This fluid is present around the joints and is produced by the synovial cells in the membrane lining of the joint spaces.It acts as a lubricant and adhesive, and provides nutrients for the avascular articular cartilage.

Large joints like knee joint contain not more than 4.0 ml of fluid. It is difficult to collect sample, unless there is an effusion.

Recommended tests on synovial fluid which give precise information of the disease are :

ROUTINE TESTS :

Gross examination (color, Clarity)

Total and differential leucocyte counts

Gram's stain and bacterial culture (aerobic and anaerobic)

Crystal examination with polarizing microscope

Other useful tests include:

Fungal and acid fast stains and culture

PCR for mycobacterial DNA

Uric acid etc.

Exercise

1. What is synovial fluid what is its function?
2. List any four investigations performed on synovial fluid?

LIST OF REFERENCES

Textbook of Medical Laboratory Technology (Second Edition) - Praful B.Godkar & Darshan P. Godkar

INTRODUCTION TO HEAMATOLOGY

- Haematology, is the branch of medicine, that is concerned with the study of blood, the blood-forming organs, and blood diseases.
- It includes the study of causes, diagnosis, treatment, out come, and prevention of blood diseases.
- Both – Production of blood and its components are affected by blood disease.

UNIT - 3

Histopathology (Lab Process)

5.0 UNIT OVERVIEW AND DESCRIPTION

- **OVERVIEW**

This unit will provide the student information about the scope of histopathology and the organizational structure of a histopathology laboratory. It will help to understand the relevant terms, procedures and working of equipments pertaining to histopathology.

Organization of a Histopathology Laboratory:

The staff to be deployedby a laboratory depends on overall work load and the different types of histopathology materials to be processed. Assuming that the laboratory consists of routine histology and special/ immunohistochemical staining sections, the employees would include a laboratory head, skilled employees in the form of histotechnicians to supervise the different sections, and unskilled employees in the form of laboratory assistants. Related areas omitted in this example should have close communications with the histology department, but maintain separate and distinct supervision.

The Chief of the Laboratory should be a trained histopathologist.

The Histotechnicians should at least have a diploma in medical laboratory technology from a recognized institution. They are responsible for specimen collection, preparation and staining.

Support Staff include attendants and clerical workers in the laboratory.

The infrastructure of the laboratory must be well designed,adequately spaced and comfortable to enable the professional and support personnel to perform their duties optimally. It must contain four defined areas:

- Reception.
- Specimen collection room.
- Processing and staining area.
- Reporting room.

- **KNOWLEDGE AND SKILL OUTCOMES**

i) To understand the scope of histopathology.

ii) To know the organizational structure of a histopathology laboratory.

iii) To know the relevant terms, procedures and working of equipments pertaining to histopathology.

- **RESOURCE MATERIALS**

1. John D. Bancroft, Marilyn Gamble. Theory and Practice of Histological Techniques. 6th ed. Churchill Livingston; Elsevier; 2008.
2. Kim S Suvarna, Christopher Layton, John D. Bancroft. Bancroft's Theory and Practice of Histological Techniques: Expert Consult: Online and Print, 7th ed. Churchill Livingston; Elsevier; 2012.
3. Derek C. ALlen, Iain R Cameron (Editors). Histopathology Specimens: Clinical, Pathological and Laboratory Aspects. 2nd ed. Springer;2012.

- **DURATION –TO BE SPECIFIED BY CBSE**

- **LEARNING OUTCOMES**

After completing this unit the students should be able to

1. Demonstrate knowledge, comprehension, and application of general techniques in the areas of:
    - Specimen accessioning.
    - Grossing methods.
    - Fixation techniques.
    - Processing of tissues.
    - Microtomy.
    - Routine and special staining.
    - Cover slipping, labeling and release of slides.
    - Storage and archiving of slides and blocks.
    - Processing of specialized tissue like bone.
2. Assemble, operate and maintain routine histology instruments.
3. Provide solutions for basic problems related to methods of general histology techniques.
4. Apply principles of lab safety in completing all laboratory work.
5. Ensure quality control while performing general histology procedures.

INTRODUCTION TO HISTOPATHOLOGY

Histopathology is a branch of pathology which deals with the tissue diagnosis of disease. A histopathological diagnosis can be made on the basis of either biopsy material taken from the patient, or from autopsy material. Tissue diagnosis helps in establishing the cause and mechanism of death by natural causes, as well as, unnatural ones (as in criminal cases or forensic pathology). It also aids in monitoring disease progression or response to treatment.

The former (biopsy material) constitutes the bulk of a histopathologist's workload. In any patient subjected to histopathological examination the final diagnosis is made by the histopathologist and this largely determines the future clinical management of the patient.

SCOPE OF HISTOPATHOLOGY

Histopathology is used for demonstration of morphological alterations in disease. The differences between diseased and normal tissues are often minor or imperceptible the majority of the methods involved in histopathology may be used for both normal and diseased tissue. The specialized methodology used for preserving and preparing tissue is known as **processing**. In order to study tissues with a microscope they must be cut into sections thin enough to be translucent. In other words tissue must be prepared in such a manner that it is one to two cells thick to enable easy visualization.

It is difficult to recognize the various components of cells and tissues on a light microscope, without **differential staining**. The stains may react chemically or physically to create as wide a variation as is possible. The staining methods can be altered to suit the needs of the examiner in order to accentuate certain tissues or organelles.

LIST OF EQUIPMENT REQUIRED FOR SETTING UP OF A HISTOPATHOLOGY LABORATORY

1. Microscope
2. Grossing station and instruments
3. Tissue processor
4. Tissue embedder
5. Microtome
6. Incubator
7. Hot air oven
8. Tissue floatation bath
9. Manual staining equipment/ Automated slide stainer
10. Refrigerator / Block cooling tray

11. Laminar flow
12. Manual immunohistochemistry staining equipment/automated immunohistochemistry stainer
13. Magnetic stirrer
14. Microwave oven
15. Mechanical/Analytical balance
16. Centrifuge
17. pH meter
18. Image analysis system

HANDLING OF THE SPECIMEN

A histopathology specimen can be transported in glass, plastic or metal containers in 10% formalin. If formalin is not readily available, one can refrigerate the specimen at 40°C to slow down autolysis. The specimen hardens after fixation so the container should be large enough for its easy removal. Fresh material is required for the following purposes:

1. Frozen section
2. Tissue culture
3. Molecular analysis and other research

STEPS IN TISSUE PROCESSING

1. Receipt of samples and identification
2. Specimen entry and acknowledgement
3. Grossing
4. Tissue processing
5. Embedding
6. Section cutting
7. Staining (Haematoxylin & Eosin and special stains)
8. Typing and dispatch of reports
9. Confidentially of reports
10. Critical test reports/ Quality control
11. Amendment of reports
12. Issue of slides
13. Review of outside slides
14. Maintenance of records/ digital archiving

Figure 5.1 : Student Microscope

5.6 ROUTINE TECHNIQUES IN HISTOPATHOLOGY

Specimen Receiving

1. All surgical pathology specimens received in the laboratory should have an accompany-ingrequisition form that gives detailed patient information and history (patient name, medical record number, age and sex, address, date and time of collection, requesting physician's name, preoperative and postoperative diagnoses, clinical history including previous biopsies, surgeries, and prior therapy).
2. The specimens are given a unique identification number that will identify each specimen for each patient.
3. Requests including photos and special stains are noted on the requisition form.

Gross Examination

Gross examination consists of describing the specimen and placing all or parts of it into small plastic cassettes which hold the tissue while it is being processed to a paraffin block. Multiple specimens from the same operative procedure are designated as letters "A", "B" etc. It is important to mention the anatomic site and provide detailed description of every specimen.

Requirements for a histopathology dissection room

1. A cutting board designed in such a fashion that the drainage of all the fluids is directed into the sink.
2. Specimen containers.
3. Shelves to accommodate the specimen containers.
4. A sink with hot and cold water.
5. Formalin.
6. Instruments including heavy and small scissors, different sized smooth and toothed forceps, a malleable probe, a scalpel handle, disposable blades, knives and pins.
7. Labels.
8. Weighing balance.

Grossing techniques

Ready to install grossing stations are now available for histopathology grossing. A pathologist, histotechnologist or biomedical scientist can gross specimens.

Key points in grossing of histopathology specimens:

- If the biopsy is very small (like a core or a mucosal biopsy, and brain biopsy) then ideally the entire tissue should be processed.

- If the specimen is very large then relevant sections should be taken by the pathologist, e.g., in a radical mastectomy specimen, relevant areas like the tumour, its margins, one or two sections from the normal looking breast parenchyma, sections from nipple, areola and lymph nodes are a must.
- Small fragments of tissue must be wrapped in thin paper.
- Even smaller fragments can be stained with haematoxylin for easy identification by the histopathologist.
- The identification of surgical margins can be facilitated by painting them with India ink or a similar dye before sectioning.

Safety precautions in the grossing room:

1. Treat all tissue/specimens with universal precautions.
2. Wear aprons/disposable gowns in the grossing laboratory.
3. Wear face masks and gloves while handling or processing specimens.
4. Ensure minimum contact with chemicals; be aware of toxicities of chemicals in the work place.
5. Clean all instruments with a disinfectant after using them.
6. Wash hands frequently.

Figure 5.2 : Gross Dissection Hoods
(courtesy Thermoshandon)

Fixation

- Tissue removed from the body undergoes a process of degeneration. This process known as **autolysis**, starts immediately after cell death and is attributed to enzymes, which cause cellular breakdown and eventual liquefaction. Organs like the liver, brain and kidney, which are rich in these enzymes undergo rapid liquefaction, whereas, tissues such as elastic fibers and collagen which have less of these enzymes, liquefy slowly. Also, inthe absence of any preservation, superimposed bacterial infection occurs to cause **putrefaction** of tissue. To avoid this, the tissue needs to be preserved or fixed.
- Fixation enables the constituents of the cells and therefore of the tissues, to be fixed in a physical and chemical state such that they can withstand subsequent treatment with various reagents with minimum loss or distortion or decomposition.

- Fixatives mostly act by denaturing or precipitating proteins, which then form a mesh work tending to hold together the other cell constituents.
- Fixation should be done immediately after removal. Amount of fixative used should be 10-20 times the volume of the specimen.

Aims of fixation:

1. Fixation helps in maintaining the spatial relationship between cells and extracellular substances, e.g., collagen, elastic fibers and amorphous ground substances.
2. It enhances the differences in refractive indices of different constituents of cells and tissues toincrease their visibility in tissue sections.
3. It renders cell constituents' insoluble, by working on tissue proteins to stabilize them.
4. It hardens the tissue and helps further handling.
5. It prevents autolysis and bacterial putrefaction (microorganisms are also composed of proteins which will be fixed and organisms killed by fixatives).
6. It enhances later staining techniques (act as mordants for certain stains, e.g., mercurials; mordants are metallic salts, which act as a bridge between the stain and tissue enabling staining to take place).

An ideal fixative should:

1. Be cheap, stable and safe to handle.
2. Prevent autolysis and bacterial decomposition.
3. Penetrate the tissue quickly and be rapid acting.
4. Preserve tissues in their most natural state and fix all its components (protein, carbohydrate, fat, etc).
5. Harden the tissue for further handling.
6. Be isotonic and confer chemical stability on the tissue.
7. Should not bind those reactive groups upon which specific staining of the tissue elements will depend.
8. Make the cellular components insoluble to liquids encountered during tissue processing.
9. Preserve tissue volume.
10. Avoid excessive hardness of the fixed tissue.
11. Enhance subsequent staining.
12. Be nontoxic and non allergenic.

Classification of fixatives:

1. Classification based on their usage:

A. Tissue fixatives

- Buffered formalin
- Buffered glutaraldehyde
- Zenker's formol saline
- Bowen's fluid

B. Cytological fixatives

- Ethanol
- Methanol
- Ether

C. Histochemical fixatives

- Formol saline
- Cold acetone
- Absolute alcohol

2. Classification based on their mechanism of action:

1. Physical methods of fixation:-

a. Heat fixation:

Heat is the simplest availablefixative. It renders each cellular component less soluble in water after fixation. Though acceptable morphological preservation can be achieved by boiling tissue in normal saline, it is only used in histopathology to accelerate other forms of fixation as well as shorten steps of tissue processing.

b. Microwave fixation:

Microwave heating can reduce time for fixation of some gross specimens from more than 12 hours to less than 25 minutes but microwaving tissue in formalin can result in production of large amount of dangerous vapors. Instead commercial glyoxal based fixatives have been introduced that do not form vapors when heated at 55°C.

c. Freeze drying and freeze substitution:

Tissues are cut into thin sections, immersed in liquid nitrogen and the water is removed in a vacuum chamber at – 40^{o}C. The tissue can be post fixed with formaldehyde vapor.

2. Chemical fixation:

Chemical fixation utilizes organic or non-organic solutions to maintain adequate morphological preservation. These are categorized into three major groups:

a. Coagulant fixatives:

Both organic and non-organic solutions coagulate proteins making them insoluble. Cellular architecture is maintained, however, because these fixatives coagulate cellular proteins leading to poor preservation of mitochondria and secretary granules, such fixatives should not be used for ultrastructural analysis.

b. Dehydrant coagulant fixatives:

The most commonly used dehydrant coagulant fixatives are alcohol (ethanol, methanol) and acetone. These fixatives disrupt the tertiary structure of proteins; change their physical properties, potentially causing insolubility and loss of function. Fixatives like trichloroacetic acid induce changes in the side chains of proteins and disrupt electrostatic and hydrogen bonds but do not precipitate proteins, they may be used in combination with other fixatives to minimize loss of nucleic acids.

c. Non-coagulant cross linking fixatives :

These fixatives form cross-links within proteins and between proteins and nucleic acids, e.g., formaldehyde, glutaraldehyde and other aldehydes like chloral hydrate and glyoxal, metal salts such as mercuric and zinc chloride and other metallic compounds such as osmium tetra oxide.

Overall, the most commonly used fixatives include:

- Aldehydes
- Mercurials
- Alcohols
- Oxidizing agents
- Picrates

Aldehydes

- Include formaldehyde (formalin) and glutaraldehyde.
- Formaldehyde is a colorless gas with a characteristic pungent odor. Commercial solution of formaldehyde in water is commonly called formalin. A saturated water solution, that contains about 40% formaldehyde by volume or 37% by mass, is called "100% formalin".
- Formaldehyde is usually used as 10% neutral buffered formalin (NBF).
- Formalin fixes tissue by forming cross-linkages (methylene bridges) in the proteins.
- A block of average tissue (4mm thick) takes 8 hrs to fix at room temperature.
- On storage commercial formalin becomes turbid specially if stored in a cold place due to formation of paraformaldehyde resulting from polymerization of formalin. This can be removed by filtration. Also all commercial solutions of formalin have 11-16% methanol which tends to inhibit formation of paraformaldehyde.
- Formalin oxidizes to formic acid which promotes autolysis, reduces the quality of staining, particularly nuclear, leaches out hemosiderin and causes precipitation of formalin (formol-heme) pigment in the tissues. To prevent this one can use formol saline with phosphates and calcium carbonate or a buffer.
- Due to the larger size of its molecule as compared to formaldehyde, Glutaraldehyde shows a slower rate of diffusion into the tissue. Consequently glutaraldehyde is not ideal for fixation of thicker tissue samples. It is mainly used for electron microscopy.

Mercurials

- Contain mercuric chloride and fix tissue by an unknown mechanism. Mercuric ions are thought to combine with acidic (COOH) or sulfur (thiol) groups of proteins. Commonly used mercurial fixatives include B-5 and Zenker's.
- Mercurials are fast and give excellent nuclear detail but penetrate relatively poorly and may cause some tissue hardness and shrinkage. They are best used for fixation of hematopoietic and reticuloendothelial tissues.

Alcohols

- The main alcoholic fixatives are methanol and ethanol. They are precipitating (or denaturing) fixatives, which act by disrupting the hydrophobic interactions in proteins that are responsible for their tertiary structure. The precipitation and aggregation of proteins is a very different process from the cross linking that occurs with the aldehyde fixatives.
- These are excellent cytological fixativesas they are quick actingand give good nuclear detail.

- They are not used as frequently as aldehydes because they cause too much brittleness and hardness of tissues.
- Examples of alcoholic fixatives include

a) Carnoy's fixative which contains 60 ml of absolute ethanol, 30 ml of chloroform, and 10 ml of glacial acetic acid, and is ideal for small tissue fragments, e.g., curettings.

b) Clarke's solution which contains 75 ml of absolute alcohol and 25 ml of glacial acetic acid, and is a good fixative for cell culture in chromosomal studies. Glacial acetic acid is not usually used independently but can be combined with the other precipitating fixatives. The alcohols, on their own can cause substantial shrinkage and hardening of tissue during fixation whereas acetic acid alone induces tissue swelling; combining the two may result in better preservation of tissue morphology.

Oxidizing agents

- Oxidizing agents include potassium permanganate, potassium dichromate, and osmium tetroxide. They cross-link proteins, but cause extensive denaturation.
- Potassium dichromate, chromic acid, and potassium permanganate are all used in certain specific histological applications.

Picrates

Picrates include fixatives containing picric acid. Examples include Bouin's fluid (picric acid, formalin and glacial acetic acid). Bouin's solution has an unknown mechanism of action; gives good nuclear detail and imparts a yellow color to all tissues, and this aids in identifying small bits e.g., testicular, gastrointestinal, and endocrine biopsies.

Specific fixatives

Fixation of selected individual tissues can be brought about by specific fixatives:

1. Eyes: Eyes are fixed in NBF. Globe must be firmly fixed in order to cut good sections for embedding.
2. Brain: The conventional fixation of whole brain takes about 2 weeks to render it firm enough to investigate the neuroanatomy and to produce sections. A perfusion technique is used which allows all the above to be accomplished and report issued in 5 to 6 days.
3. Lungs and GIT: Typically fixed in NBF.
4. Testes: These are routinely fixed in NBF. Bouin's fixative is used for testicular biopsies.
5. Spleen and blood filled cavities: Fixed in Zenker's fluid.
6. Lymph node: Fixed in B5.

7. Renal biopsies :
   i. NBF for routine histology.
   ii. Carson's modified Millonig's fixative or 2% buffered glutaraldehyde (pH713) for ultrastructural analysis.
   iii. Commercial transfer solutions, e.g., Zeus for immunofluorescence examination.

Useful formulae for fixatives:

1. Formol Saline

Formaldehyde in isotonic saline can be used for routine histopathology fixation. This mixture (formol saline) was extensively used as a fixative prior to the introduction of phosphate buffered formalin. The former, under acidic conditions, often produces formalin pigment which can be prevented by using buffered formalin.

Constituents:

- 40% formaldehyde: 100 ml
- Sodium chloride: 9 g
- Distilled water: 900 ml

Fixation time: 12 – 24 hours

2. 10% Neutral Buffered Formalin or NBF

This fixative is the most widely used formaldehyde-based fixative for routine histopathology. It is preferred to formol-saline as the buffer prevents the formation of formalin pigment.

Constituents:

Tap water	900 ml
Formalin (37% formaldehyde solution)	100 ml
Sodium phosphate, monobasic	4g
Sodium phosphate, dibasic anhydrous	6.5 g

pH should be 7.2 – 7.4.

Biopsies require a fixation time of 1-4 hours minimum; a longer time may be required for larger specimens.

3. 10% Formol Calcium

This is a good fixative for preservation of lipids. It can be prepared in two different ways:

a) 10% Formol Calcium – Baker

Constituents:

Formaldehyde (37%) – 100ml

Tap water - 900ml

Calcium chloride – 20g

b) 10% Formol calcium – Lillie

Constituents:

Formaldehyde (37%) – 100ml

Tap water- 900ml

Calcium acetate – 20g

This fixative should be applied at least for overnight, but fixation time may extend to a few days.

4. Mercurial fixatives (Fixation time is 12 to 24 hrs)

a) Zenker's solution

Mercuric chloride precipitates protein and rapidly penetrates the tissue. Zenker's fixative is excellent for nuclear detail and connective tissue staining but lyses red blood cells due to the presence of acetic acid and may also cause excessive hardening. It is recommended for blood containing specimens and considered a good fixative for PTAH and trichrome staining however may produce mercury and chrome pigments which should be removed from sections prior to staining. After water washing, storage of tissue in 70% ethanol is recommended.

Constituents:

Distilled water	250 ml
Mercuric chloride	12.5 g
Potassium dichromate	6.3 g
Sodium sulfate	2.5 g

Add 5ml of glacial acetic acid to 95 ml of Zenker's solution before use.

b) Helly's fluid

It is considered ideal for bone marrow and cardiac muscle. As with Zenker's fixative it may produce mercury pigment (to be removed from sections prior to staining), chrome pigment

(forms if tissue is not washed in water prior to processing) and because of the low pH of this fixative formalin pigment formation may also occur. After water washing, tissue should be stored in 70% ethanol.

Constituents:

Stock solution:

Potassium dichromate - 25g

Mercuric chloride - 50g

Sodium sulphate - 10g

Distilled water - 1000ml

Working solution:

Stock solution - 100ml

Formalin (40% aqueous solution of formaldehyde) - 5ml

The fixative solution should be prepared just before use. Fixed tissue must be washed for 24 hours in running tap water prior to processing.

c) Schaudinn's solution (Fixation time 20-30 minutes)

Schaudinn's solution is used for wet smears in cytology. It is unsuitable for tissue, being not much better than a simple solution of mercuric chloride. Post fixation mercury pigment needs to be removed. Due to its mercuric chloride content this fixative may corrode metal.

Constituents:

Distilled water - 50ml

Mercuric chloride – 3.5g

Absolute ethanol – 25ml

d) B5 (Fixation time: 4 - 8 hours)

It gives excellent nuclear detail and is the recommended fixative for special stains and immunohistochemistry (IHC). Sections will require the removal of mercury pigment prior to staining. Despite its mercury content and consequent problems with disposal this fixative is popular for fixation of hematopoietic and lymphoid tissue.

Constituents:

Stock solution

- Mercuric chloride: 12 g
- Sodium acetate anhydrous: 2.5 g
- Distilled water: 200 ml

Working solution (should be prepared immediately before use)

- B-5 stock solution: 20 ml
- 40% formaldehyde: 2 ml

5. Bouin's fluid

It is prepared with saturated picric acid, formaldehyde and acetic acid and is an excellent fixative for preserving soft and delicate structures like renal, gastro-intestinal and testicular biopsies and endocrine gland tissue. It is also used as a mordant in various trichrome procedures. It preserves glycogen well but usually lyses erythrocytes. It stains tissue bright yellow due to picric acid. Excess picric should be washed from tissues prior to staining with 70% ethanol.

a) Alcoholic Bouin's fluid (for renal biopsies)

Constituents:

80% alcohol	-	80	ml
Concentrated formalin	-	1.5	ml
Glacial acetic acid	-	5.0	ml
Saturated picric acid	-	2.0	gm

b) Regular Bouin's fluid (for testicular biopsies)

Constituents:

Saturated picric acid	-	750	ml
Concentrated formalin	-	250	ml
Glacial acetic acid	-	50	ml

6. Clarke's solution (fixation time: 3 - 4 hours)

It is mainly used on frozen sections and smears as it has a very short fixation time. It gives

good nuclear preservationbut lipids are extracted. Tissues can be transferred directly into 95% ethanol.

Constituents:

- Ethanol (absolute): 75 ml
- Glacial acetic acid: 25 ml

7. Formol acetic alcohol (Fixation time 1 minute).

Used for cytology smears, cytospin preparations or frozen sections. The sections should be washed in water before staining.

Constituents:

95% methanol - 100ml

Glacial acetic acid - 3ml

40% formaldehyde: 10 ml

8. Carnoy's fluid (Fixation time 1-3 hours)

It penetrates rapidly and gives good preservation of nuclei and glycogen. It lyses erythrocytes and dissolves lipids and can produce excessive hardening and shrinkage. Fixed tissue should be processed immediately or transferred to 80% alcohol.

Constituents:

Ethanol - 60ml

Chloroform - 30ml

Glacial acetic acid - 10ml

9. Formol sublimate (Fixation time 4-6 hours)

May cause excessive hardening and shrinkage but gives excellent results with acid dyes and metachromatic stains.

Constituents:

Formalin (40% aqueous solution of formaldehyde) - 100ml

Mercuric chloride (saturated aqueous) - 900ml

Fixed tissue should be transferred to 80% alcohol.

10. Michel's fixative for immunofluorescence (Fixation time 24-48 hours)

Constituents:

0.81g potassium citrate

0.0625g N-ethylmaleimide – To be handled with care

0.123g magnesium sulphate

100mls distilled water

Before use add 55g ammonium sulphate and allow it to dissolve.

Adjust pH to 7.0-7.2 with 1M KOH.

Place tissue biopsies in fixative for 24-48 hours. Wash tissues in buffer, three times over 10 minutes, and freeze at -70oC.

Removal of fixation induced pigments:

1. Formalin pigment

1. Bring sections to water (dewax the sections, rinse them in 100% alcohol, 70% alcohol, followed by distilled water).
2. Treat in saturated alcoholic picric acid for 30 minutes to 2 hours.
3. Wash well in running tap water.
4. If yellow staining of the section persists rinse in dilute lithium carbonate.
5. Rinse in tap water.

2. Mercury pigment

1. Brings sections to water.
2. Treat in Lugol's iodine for 2 minutes.
3. Decolorize in 5% sodium thiosulphate for 5 minutes.
4. Wash well in running tap water.

3. Dichromate pigment

1. Bring sections to water.
2. Treat in 2% HCl in 70% alcohol for up to24 hours.
3. Rinse in tap water.

Mounting of museum specimens

- Prior to mounting, specimens should be trimmed according to specifications and fixed in fixatives to avoid decomposition or distortion.

- The volume of the fixative should be 10 times the volume of the specimen.
- Specimens should be suspended in the fixative and contact with the glass/ perspex container should be avoided.
- Penetration rate of the fixative into some organs such as liver, kidney, and spleen is very slow. This can be overcome by direct injection of the fixative.
- 10% formalin can be used; however, modified solutions containing additives should be used to improve the life of the specimens displayed.
- Examples of some of the methods are Romhanyi's Method, Wenthworth's Method, and Kaiserling's Method.
- Specimen fixation for museum mounting is based on a formalin fixation technique derived by Kaiserling (1897). According to Kaiserling the initial fixation should be in a neutral buffered formalin solution and the specimen should be transferred for final preservation to a glycerin solution to allow long term display. This method helps in color preservation.

a. Kaiserling I Solution:

Formalin 2 liters

Potassium acetate 425 g

Potassium nitrate 225 g

Distilled water to make up to 20 liters

* Specimen is stored in the solution for 1 month depending on the size of the specimen.

b. Kaiserling II Solution:

Alcohol 95%

* Store specimen in this solution for 10 minutes to 1 hour depending on size of specimen.

c. Rejuvenator Solution:

Pyridine 100 ml

Sodium hydrosulphite 100 gm

Distilled water 4 liters

* Formalin decreases the natural color of the specimen. However, the rejuvenator solution restores the color.

a. Kaiserling III Solution:

Potassium acetate 300 g

Glycerine 6 liters

Distilled water to make up to 10 liters

Thymol crystals added to prevent moulds.

Leave solution to stand for 2 – 3 days before using to ensure proper mixing of chemicals. Add 1% pyridine as stabilizer. This solution acts as permanent fixative. This solution easily turns yellowish and needs to be replaced to restore color of the specimen.

Tissue processing

The technique of getting fixed tissue into paraffin in order to make thin microscopic sections of it is called tissue processing. It can be achieved manually or by automation. The stages involved in processing are the following:

1. Dehydration
2. Clearing
3. Impregnation.
4. Embedding

Factors affecting the rate of processing include:

1. Agitation :-

Using manual processing method, agitation is difficult to achieve and would be very tissue consuming. Automatic tissue processing methods incorporate agitation using rotation or virtual oscillation of the tissue basket as in the case of carrousel type machines or a pumping system. The rate of agitation should not be too slow so that it is effective but neither should it be violent as this can cause small fragments of tissue to be damaged. Effective agitation may reduce overall processing time by up to 30%.

2. Heat :-

Heat increases the rate of penetration and the interchange, but as most of the processing fluids are highly inflammable this could be dangerous. Enclosed processors are designed to be safe when heat is used. Temperature limited to 45°C can be used effectively.

3. Viscosity :-

Most fluids used during standard dehydration and clearing have similar viscosities.

4. Vacuum :-

Incorporation of vacuum systems to reduce pressure in enclosed processing machines with dense and fatty tissues such as brain greatly reduces the impregnation time.

Dehydration:

- The main steps in this process are dehydration and clearing. Tissues are routinely embedded in paraffin as the latter is similar in density to the tissue and can be sectioned at anywhere from 3 to 10 microns (usually 6-8 microns). Tissue fixed in aqueous solutions cannot be directly infiltrated with paraffin. First, the water from the tissues must be removed by dehydration. This is usually done with a series of graded alcohols; 70% to 95% to 100%.
- Many dehydrants can be used but have some disadvantages, e.g., acetone (is inflammable), and dioxane (has toxic fumes). Other dehydrating fluids include industrial methylated spirit (denatured alcohol) which contains ethanol to which a small amount of methanol is added. Isoprophyl alcohol is recommended while using a microwave oven. Acetone is rapid in action but has poor penetration and causes brittleness in tissues in case of prolonged use.

Clearing

The next step consists of replacement of the dehydrating solution with a substance that will be miscible with the embedding medium or paraffin. During dehydration, water in tissue is replaced by alcohol. As paraffin wax is not alcohol soluble, alcohol needs to be replaced with a substance in which wax is soluble. The commonest clearing agent is xylene. Other clearing agents include chloroform, benzene, carbon tetrachloride and toluene. Toluene is good but expensive. Chloroform can be a health hazard, and is slow. Newer clearing agents available include limolene, a volatile oil found in citrus peels and clearite, a long chain aliphatic hydrocarbon compound.

An ideal clearing agent should have the following properties:

- Rapid removal of the dehydrating agent.
- Easy removal by molten paraffin wax.
- Minimal tissue damage.
- Less flammability and toxicity.
- Cost effectiveness.

Prolonged exposure to most clearing agents causes the tissue to become brittle and therefore more difficult to section.

Embedding

- Impregnation is process of replacing the clearing agent with the embedding medium. Paraffin wax continues to be the most popular embedding medium for histology for a

number of reasons. It is cheap, easily handled and section production provides few difficulties. Certain additives are added in paraffin wax like lemon, rubber, diethylene glycol, ceresin either alone or in combination to increase the hardness of paraffin wax, thus enabling then sections to be cut.

- Paraffins can have different melting points, for obtaining different degrees of hardness.
- Embedding is undertaken at melting point temperature of paraffin wax, which is 54-60^{o}C. Volume of wax should be about 25-30 times the volume of tissues. Application of vacuum within the tissue processor aids in the penetration of the embedding agent.

Types of moulds

a) Leuckhart's L pieces - These are two 'L' shaped pieces of metal usually brass, which are resting on a flat metal or glass plate. The L shaped pieces can be adjusted to modify the mould depending on the size and shape of the tissue.

b) Compound embedding units - consists of square shaped brass or metal plates in a series of interlocking plates.

c) Others like plastic embedding blocks (Tissue Tek system)

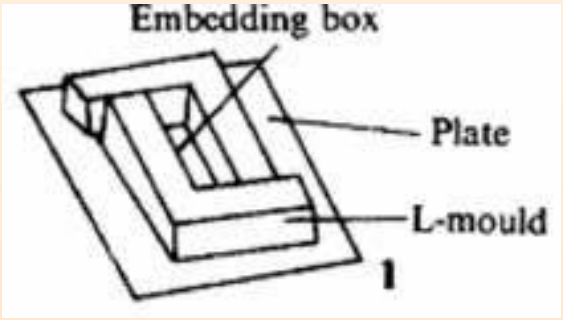


Figure 5.3 : L-moulds or embedding box

Techniques of casting

1. Molten paraffin wax is heated to a temperature 2-3^{o} above the melting point and is then poured into the mould so as to cover the thickest tissue block completely.
2. The tissue is then transferred to the mould with prewarmed forceps (to prevent the wax to stick to it). The tissue is gently placed in the wax to orient it in such a way that the surface to be sectioned is facedown. Care should be taken that there are no entrapped air bubbles.


Figure 5.3 : Plastic embedding blocks (tissue Tek system)

3. The label is fixed in position by pressing one edge against the side of the solidifying wax in the mould.

4. When a film of solid wax is formed on the surface, of the whole block, the mould is submerged in cold water at 20°C or transferred to a refrigerated surface.
5. When the blocks undergo complete solidification they are removed from the mould. The tissue surface facing the mould base is from where the sections are to be cut and this surface needs to be trimmed lightly with a scalpel so as to expose the tissue. Trimming is an important step for proper alignment and orientation of the specimen.

   Alternatives to paraffin embedding include various plastics that allow thinner sections, e.g., methyl methacrylate, glycol methacrylate, araldite, etc. Plastics require special reagents for dehydration and clearing that are expensive.

Technical points to be taken care off during casting

- The cutting surface of the tissue should be facing down towards the bottom of the mould.
- If more than one tissue has to be casted they should be kept at the same depth.
- It is to be ensured that all four corners of the block are in one horizontal plane.
- The tissue should have at least 2 mm wax around its edges.
- Mineral or machine oil can be smeared on the inner surface of the mould to enable easy removal of block.
- There should be no whitish discoloration around the tissue in the block as this indicates crystallization. Crystallization is due to moisture or due to incomplete removal of clearing agent.
- Most tissue sections are cut from the largest area but some tissue needs special mention.
- Tissue of tubular nature is cut transversely so should be embedded vertically.
- Skin is cut in a plane at right angles to the surface so should be embedded at right angles to the bottom.
- Muscle biopsy should be sectioned in both transverse and longitudinal planes.

Alternative embedding media

- These media are used when processing agents remove or destroy tissue components that are the object of investigation.
- Sections are required to be thinner.
- The use of heat may adversely affect tissue.

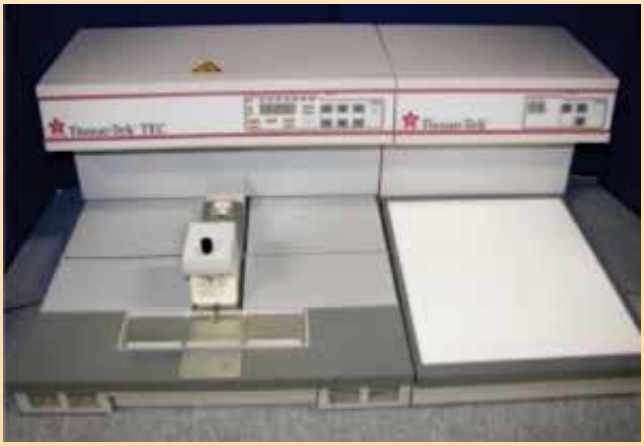


Figure 5.5: Embedding System (courtesy Thermoshandon)

- The impregnation medium is not sufficiently hard to support the tissue.

  Alternative embedding media include -

  1. Resin – Embedding media for undecalcified bone.
  2. Agar – Cohesive agent for small friable pieces of tissue after fixation.
  3. Gelatin – For frozen sectioning.
  4. Celloidin – Use in neuropathology

Automated tissue processor

An automated tissue processor has the following advantages:

1. The tissue can be mechanically transferred from one reagent to another both during day and night.
2. The overall processing time is reduced courtesy the continuous agitation in automated processor.
3. Automation reduces the possibility of human errors of leaving the tissue for long time in one solution.

Tissues are usually more rapidly processed by machine than by manual methods. Similar to manual processing the tissue is most conveniently processed through stages of dehydration, clearing and infiltration automatically by machine. There are two broad types of automatic tissue processors - tissue-transfer and fluid-transfer types.

Tissue-transfer processors: In this type of a processor, the tissue contained in a basket is transferred through a series of stationary reagents arranged in-line or in a circular carousel plan. The rotary or carousel is the most common model of automatic tissue processor. Agitation is achieved either by vertical oscillation or rotary motion of the tissue basket. Processing schedules may be card-notched, pin or touch pad programmed.

In fluid-transfer processors, the tissue remains stationary and the processing fluids are pumped to and from a closed chamber Fluid-transfer processors are designed to overcome the drawbacks of the tissue-transfer machines. The sealed chambers prevent the tissues from spillage, drying out and the fumes and reagent vapors are vented through filters or retained in a closed-loop system.

Tissue transfer machines can be of two types, the traditional ‘carousel’ type and the “enclosed pump fluid” type. Both usually have the facility for 12 separate stages in processing. Enclosed processors have an advantage over the carousel type in that if there is any fault with the electronics or mechanics then the machine stops and sounds an alarm. Also, these have more rapid schedules, less fluid spillage, and containment of toxic fumes emitted. In most laboratories an overnight schedule is used, of approximately 16-18 hours duration.

The various parts of an automated tissue processor are

(a) Tissue containers - These are also called "cassettes". The tissue to be processed is placed in an appropriate sized, closed cassette, together with a label. These are then placed in the tissue basket within which they undergo all the stages of processing.

(b) Beakers and wax baths - Most machines are equipped with ten beakers and 2 wax baths thermostatically controlled at 56°C + 4°C. The beakers are filled with appropriate reagent fluids and the wax baths are filled with wax which is kept in the molten state.

(c) Stirring mechanism - The basket is attached to the arms of the processor, one of which is designed in such a manner so as to bring about the rotation of the basket nearly at the rate of one revolution per minute.

(d) Timing mechanism – The timer is meant to ensure that the tissue is kept in different reagents and wax for an optimum time. The time schedule for processing has to be strictly adhered to as otherwise the tissue will not be optimally processed.

Technical points to be noted

1. Fluid and wax beakers must be filled up to the appropriate mark.
2. Any spillage of the fluid should be wiped away.
3. Accumulations of wax must be removed from beaker, covers, lids and surrounding areas.
4. Wax bath thermostats should be set at satisfactory levels usually 2-3°C above the melting point of wax.
5. Particular attention should be paid to fastening the processing baskets on the carousel type of machines.
6. Timer should be set with utmost care when loading the machine.
7. Paraffin wax baths should be checked to ensure that the wax is molten.

Automated processing schedule

1. 80% alcohol (holding point) 1 hour
2. 95% alcohol 2 hours
3. 95% alcohol 1 hour
4. 100% alcohol 1 hour
5. 100% alcohol 1 hour

Figure 5.6: Excelsior Tissue Processor (courtesy Thermoshandon)

6. 100% alcohol 1 hour
7. Chloroform 1 hour
8. Chloroform 1 hour
9. Chloroform 1 hour
10. Paraffin wax 2 hours
11. Paraffin wax 2 hours
12. Paraffin wax 2 hours

Note: Frequent filtration and changes of solution are needed

Sectioning

Paraffin section cutting requires the following equipment:

1. Floatation (water) bath
2. Slide drying oven or hot plate
3. Fine pointed or curved forceps
4. Stable one camel haired brush
5. Scalpel
6. Slide rack
7. Clean slides
8. Teasing needle
9. Ice tray
10. Chemical resistant pencil.

Once the tissue has been embedded, it must be cut into sections with a thickness of 3-5µm (ensuring that only a single layer of cells makes up the section). This is achieved with an instrument called a "microtome".

Microtomy

Microtomy is the means by which tissue can be sectioned and attached to a surface so that examination by microscopy can take place. Sectioning paraffin wax embedded tissue blocks is the commonest way of achieving this. The basic instrument used in microtomy is a microtome.

Types of microtome

1. Hand microtome (used in botanical sections).
2. Rocking microtome (used for sectioning animal tissues of soft nature).

3. Rotary microtome (most frequently used microtome; can cut a large number of sections at a time, particularly serial sections). The basic mechanism of this microtome requires the rotation of a fine advance hand wheel through 36° moving the specimen vertically past the cutting surface and returning it to the starting position. The rotary microtome may be manual (completely manipulated by the operator), semi- automated (one motor to advance either the fine or coarse hand wheel) or fully automated (two motors that drive both the fine and the coarse advance hand wheel). Advantages include – the ability to cut thin 2-3 mm sections and easy adaptation to all type of tissue, that is, hardy fragile, fatty sectioning.

Figure 5.7: Leica RM 2135 Rotary Microtome

4. Freezing microtome (have a non-movable tissue block with the cutting action being the motion of the knife).
5. Base sledge microtome (can be used for sectioning of tissues of all types, sizes and degree of hardness).
6. Vibrating knife microtome (can cut sections without tissue fixation, impregnation or freezing; used for animal and botanical studies).

Knives: These are either made of standard steel, glass or diamond or thin disposable variety. Knives are classified by the manner in which they are ground and seen in their cross section.

Types of knives:

- Plane wedge: It is used for wax embedded tissue with any microtome, synthetic resin embedded tisuue as well as frozen sectioning.
- Plano concave: Used for celloidin section since the blade is thin and will vibrate when for used for other harder materials.
- Biconcave: It is recommended for paraffin section cutting on rocking and sledge type of microtome.

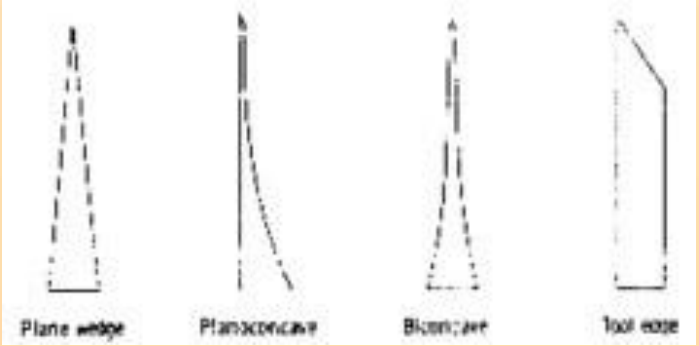


Figure 5.8: Types of microtome knives

- Tool edge: This is used for cutting very hard tissues like undecalcified bone.

Waterbath: Itisathermostatically controlled container for paraffin wax of melting point 56°C. It is painted black from inside for easy visibility of sections. Distilled water should be used; a water temperature of 45°C is sufficient; addition of a trace of detergent to water is beneficial in flattening of sections.

Figure 5.9: Standard Tissue Bath

Hot plate or drying oven: Drying of sections is achieved at around the melting point of wax.

Brush, seeker, and forceps: It is required for removal of folds and creases in sections after floating out.

Slides: The most commonly used slide size is 76 x 25 x 12 mm as most sections fit on to this size.

Figure 5.10: Large Slide Warmer

Diamond pencil: This is needed to write the identification details like name or acquisition number.

Section adhesives: An adhesive (albumin, gelatin, araldite, starch, resin, and poly L lysine), is a substance which is smeared on to the slides so that the sections stick well to them. Tissue sections which are adequately thin and thoroughly dried generally do not require an adhesive, as in case of routine H&E staining, but for histochemical and immunological methods using alkaline solutions e.g., ammonia, which tend to remove sections from slide, adhesives may be required. Adhesive is also required for blood clots, and decalcified tissues which have a tendency to detach themselves from the slide.

Steps in cutting of paraffin embedded tissue:

Vertical sectioning is cutting perpendicular (i.e. at right-angles) to the surface of the tissue. This is the most common method. Horizontal sectioning is often done for the study of hair follicles and structures that include hairs, hair follicles, erector pili muscles, and sebaceous glands. These structures are collectively called "pilosebaceous units".

Steps

1. Blocks are chilled on a refrigerated plate or ice tray for 10 minutes before sectioning.
2. Insert the appropriate knife/blade in the knife/blade holder and screw it tightly in position.
3. Blade clearance angle is adjusted (clearance angle prevents contact between the knife facet and the face of the block; the facet angle is the angle between the two facets that form the cutting edge. For routine use knives and disposable blades are made with a facet angle of approximately 35°.The clearance angle should be set at 3-4 degrees and angle of slope should be set permanently at 90 degrees).
4. The block is wiped dry and clamped in the block holder on the microtome in such a position that the surface of the block just touches the knife edge.
5. Next step is trimming for which the block is moved forward so that it is almost touching the knife. To trim away any extra wax and to expose a suitable area of tissue for sectioning, the section thickness is adjusted at 15 microns.
6. Section thickness is set at 4-6 microns. The microtome is now moved rhythmically with the right hand operating the microtome and left hand holding the sections away from the knife. A ribbon is formed due to the slight heat generated during cutting, which causes the edges of the sections to adhere.
7. Once cut, the tissue sections/ribbons are carefully transferred to a thermostatically controlled warm water bath (maintained at a temperature 4-6ºC below the melting point of paraffin wax). Here they are allowed to float on the surface and then scooped up onto a slide. Slides should be clearly labeled, and then allowed to dry upright at 37ºC for a few hours to gently melt the excess paraffin wax, leaving the tissue section intact.

   It is important to have a properly fixed and embedded block to prevent artifacts during sectioning. Common artifacts include tearing, serrations, holes, folding, etc.

Bone techniques

Techniques for demonstration of bone and its components are possibly more varied and difficult than for any other tissue. For bone biopsies, decalcification is needed in order to produce paraffin sections. Iliac crest trephine biopsies can be bisected longitudinally, half for

decalcification and paraffin techniques, and half for undecalcified bone Methylmethacrylate (MMA) sections.

Decalcification:

In order to obtain satisfactory paraffin or celloidin sections of bone, inorganic calcium must be removed from the organic collagen matrix, calcified cartilage and surrounding tissues. This is called as decalcification and is carried out by chemical agents, either with acids to form soluble calcium salts or with chelating agents that bind to calcium.

Specimen Selection:

Specimen should ideally be 4 to 5 mm in thickness; overall size and volume is less critical as the thickness of the sample. If bone slabs are too thick, both decalcification and processing are prolonged.

Types of Bone:

- **Cortical:** Dense compact bone which takes a long time to decalcify. Sections from this type of bone appear solid except for microscopic spaces (e.g., skull, pelvis, and mandible).
- **Cancellous:** Spongy bone which consists of thin branching trabeculae, which merge to form a meshwork creating intertrabecular spaces filled with marrow (e.g., tibia, femur, etc.). Takes less time to decalcify.
- **Marrow:** Requires a very short time for decalcification (1hr).

Fixation:

Fixation prior to decalcification is critical since the acid in the decalcifying solution will destroy the cellular morphology of unfixed tissue. Types of Fixatives used with decalcification include:

1. 10% NBF (neutral buffered formalin):

   Suggested fixation times:

   - Debridements, bone chips, fragments: 24 hours
   - 6 mm osteonecrosis bone cores: minimum 48-72 hours
   - 5 mm slab sections of tumors, femoral heads, knees: 4-7 days

2. B5:

   - Excellent preservation of nuclear detail.

- Suggested fixation time for bone marrow cores is 1 hour.

Decalcifying solutions

Commonly used decalcifying solutions are of the following types:

1. Strong acids include nitric and hydrochloric acids. They maybe used as simple aqueous solutions with a recommended concentration of 5-10%. They decalcify rapidly, may cause tissue swelling and can seriously damage tissue stainability if used longer than 24-48 hours. Strong acids can be used for needle and small biopsy specimens to permit rapid diagnosis within 24 hours or less. They can also be used for large or heavily mineralized cortical bone specimens with carefully monitoring of the decalcification progress by a decalcification end point test.

a) Nitric acid

Constituents:

- Concentrated HNO3 - 5-10ml
- Distilled water- up to 100ml

Advantages:

- Rapidly acting (decalcification time is less than 4 hours for most calcified specimens).
- Gives a brilliant staining reaction

Disadvantages:

- Nitric acid causes yellow discoloration of the tissue which will interfere with subsequent staining reaction. This can be prevented by adding 1% urea to pure nitric acid.
- May damage cellular morphology. Mineral acid decalcifiers are not recommended for delicate tissues such as bone marrow.

b) Hydrochloric acid

This renders nucleic acids less susceptible to hydrolysis when combined with B-5 (used for bone marrow).

Advantage:

- Acts rapidly (4 hours or less for most calcified specimens)

Disadvantages:

- Requires thorough fixation of tissues before decalcification.

- The tissue should be thoroughly washed between formalin fixation and HCl as formalin and hydrochloric acid combined together give off a carcinogenic gas, namely bischloromethyl ether.
- Overdecalcification results in poor histological detail (swelling, fragmentation, and poor nuclear staining).
- HCL can irreversibly corrode aluminum, nickel and even stainless steel equipment.

2. Weak, organic acids include formic and acetic acid. Formic acid is the only weak acid used extensively as a primary decalcifier. Formic acid can be used as an aqueous (5-10%) buffered solution in combination with formalin. The formalin (10%) formic acid mixture simultaneously fixes and decalcifies and is recommended for very small bone pieces or Jamshidi needle biopsies. It is also suitable for most routine surgical specimens particularly when immunohistochemical (IHC) staining is needed.

Advantages:

- Post decalcification staining is sharper including IHC.
- Both fixation and decalcification can occur at the same time as the decalcifying solution contains formalin. Safer to use than stronger acids
- Does not impair nuclear staining

Disadvantage:

Slower than strong acids

3. **Chelating agents**

The primary chelating agent used for decalcification is ethylenediamine tetra acetate (EDTA) which binds the ionized calcium.

Constituents:

EDTA disodium salt - 5.5g

Distilled water - 90ml

Formalin - 10ml

Advantages:

1. This is a gentle decalcifying agent that does not damage tissues or their stainability.
2. It is an excellent bone decalcifier for IHC enzyme staining and electron microscopy.

Disadvantages:

- It is slow acting. Calcium is removed by chelation and the process may take from 1-8 weeks at room temperature depending on the size of the sample.
- Judging when the decalcification is complete is more difficult than with acid solutions as the ammonium oxalate test cannot be used. X-raying the sample is the best way to ensure that all traces of calcium have been removed.

4. Ion exchange resins with acid decalcifying solutions can be used for a quick and reliable decalcification. The resins commonly used are ammonium forms of suphonated polystyrene resins. The volume of fluid will be 20-30 times the bulk of the specimen. Formic acid containing decalcifying fluid will be better results. After use with resins the tissue must be washed twice in diluted HCL and followed by washing in running tap water for 3 times

5. Electrolytic decalcification aids in speedy decalcification without any damage to morphology but excessive heat produced may cause charring of the specimen if care is not taken. These are organic compounds having the capacity to bind with calcium metals. Tissue decalcified by this method shows minimal artifacts and good staining results

Factors affecting the rate of decalcification

1. Concentration of decalcifying agent: Generally more concentrated acid solutions decalcify bone more rapidly but are also more harmful to the tissues. A combination of fixative acid decalcifying solution is advantageous as the decalcification rate cannot exceed the fixation rate or the acid will damage or macerate the tissue before fixation is complete. Ideally, acid solutions should be end point tested and changes should be made daily to ensure that tissues are not left in acids too long or over decalcified.
2. Temperature: The optimal temperature for acid decalcification is 25^{o}C.
3. Agitation: It speeds up decalcification.
4. Suspension: The decalcifying agent should make contact with all surfaces of a specimen.

Tips for proper decalcification

- Specimens should be decalcified in acid solutions 20 times their volume.
- Specimens must be fixed before exposure to an acid solution. If a combination of fixative and decalcifying solution is used, the specimen should be at least partially fixed first.

- Fresh tissue is fixed in 10% buffered formalin for 12-48 hours depending on the size and density of the bone. Tissue to be decalcified ideally should not exceed 4-5 mm in thickness but larger pieces may require partial decalcification with further subsequent sectioning. A fine- toothed hacksaw or the Stryker saw may be used to obtain thin pieces of bone.
- The decalcifying solution should be changed every day until decalcification is complete. Decalcification time varies from 24 hours to days or months depending on size of the specimens. On completion of the decalcification the specimen is rinsed in water briefly and transferred to ammonia solution to neutralize acids left in specimens for 30 minutes.
- Fixed specimens are washed in slowly running tap water for a minimum of 30 minutes. Larger specimens are washed up to a maximum of 1 hour. Avoid rinsing in rapidly running water.
- The specimen should be suspended so it is not in contact with any of the surfaces of the container. This ensures exposure to all specimen surfaces and enables the precipitated calcium salts to sink to the bottom of the container.
- Embed the harder cortical bone so that it is the last surface to be sectioned and at an angle so that the knife does not contact the entire surface at once.
- A heavy duty knife or blade might be necessary for section cutting.

  Difficult specimens can be soaked in ice water to ease cutting.

An ideal decalcifying agent

- Can completely remove calcium salts from tissues.
- Causes minimal cell/tissue destruction or distortion of components.
- Does not adversely interfere with the subsequent staining.

Completion of decalcification

Ideally bone should be taken out of the acid solution as soon as all the calcium has been removed from the bone and this requires frequent monitoring. So it is important for a laboratory to control the decalcification procedure by using decalcification end point tests.

These include

1. X-ray (the most accurate way) but not always feasible.
2. Physical tests: The physical tests include bending the specimen or inserting a needle, pin, razor, or scalpel directly into the tissue. The latter can cause tears and artifacts in

the specimen. Bending of the specimen is less disruptive but not conclusiveregarding removal of all calcium salts. Check the rigidity of the specimen and wash thoroughly prior to processing.

3. Chemical tests: Chemical test methods include the calcium oxalate test. This method involves detection of calcium in acid solutions by precipitation of insoluble calcium hydroxide or calcium oxalate solutions. The following solutions are needed to chemically test for residual calcium.

5% Ammonium Hydroxide Stock:

Ammonium hydroxide, 28% ---------- 5 ml

Distilled water ---------- 95 ml

Mix well

5% Ammonium Oxalate Stock:

Ammonium oxalate---------- 5 ml

Distilled water ---------- 95 ml

Mix well

Ammonium Hydroxide/Ammonium Oxalate Working Solution:

Use equal parts of the 5% ammonium hydroxide solution and the 5% ammonium oxalate solution.

Testing for remaining calcium in decalcifying tissue:

1. Take approximately 3ml of decalcification solution from the tissue container and place it in a test tube.
2. Test with a piece of litmus paper and neutralize the fluid by adding strong ammonia solution drop by drop until the paper just turns blue. If the litmus turns too blue add more decalcification fluid and try again.
3. Add an equal quantity (3ml) of saturated ammonium oxalate shake the tube and leave it for ten minutes.
4. If after ten minutes there is any precipitate at all (indicated by cloudiness) calcium is still present and the specimen needs further decalcification.

Staining

The embedding process has to be reversed to get the paraffin wax out of the tissue and allow water soluble dyes to penetrate the sections. Therefore, prior to any staining, the slides are "deparaffinized" by running them through xylene to graded alcohols to water. All stains have a specific methodology. Staining can be done either manually or in an automatic stainer.

1. Manual Staining:

In a small laboratory with less work load, this is the method of choice. Although it is time consuming, it is cost effective. Different reagent containers are placed serially based on the prescribed methodology and the slides are moved from one container to another manually.

2. Automated staining:

In a histopathology laboratory with a large work load, an automatic stainer is required. This has containers of staining reagents which are arranged sequentially based on the recommended method and a timer, to control the duration for which the slides remain in a given container. It has a mechanical device which shifts the slides from one container to next after the specified time.

Advantages of automated stainers:

a. They reduce the man power required
b. They control the timing of staining accurately
c. Large number of slides can be stained simultaneously
d. Amount of reagents used is less.

Classification of Dyes:

1. According to source
    a. Natural, e.g., hematoxylin, carmine, orcein and saffron.
    b. Synthetic, e.g., aniline dyes
2. Based on Tissue affinity
    a. Acidophilic (cationic dyes and stain nuclei, basophilic granules or bacteria).
    b. Basophilic (anionic dyes and stain mainly cytoplasm, eosinophilic granules).
3. Based on Chemical Structure
    a. Azo-dyes
    b. Thiazines

c. Xanthene dyes

d. Triphenyl and diphenyl methanes

e. Acridine dyes

Tissues get stained due to

a) Physical properties as in simple solubility e.g. fat stains; or adsorption (property by which a large body attracts to itself minute particles from a surrounding medium).

b) Chemical reactions as in the case of acidic and basic dyes (acid dyes stain basic elements and basic dyes stain acidophilic material) are also responsible for staining.

The basic steps in staining and mounting paraffin sections are as follows:

1. Deparaffinization

   It is essential to remove the wax from the section to allow staining. This is usually done by xylene. A properly dewaxed section should appear clear and transparent.

2. Hydration

   Most of the stains used are aqueous or dilute alcoholic solutions, therefore the sections have to be brought to water before the stains are applied. This is achieved with graded alcohols. First change is made to absolute alcohol or acetone followed by 90%, 70% alcohol and finally water. The sections should now appear opaque.

3. Staining

   The most common stain applied for histological study is Haemotoxylin and Eosin.

   Special stains may be used as and when indicated.

4. Washing and rinsing of tissue sections

   This eliminates carrying over of one dye solution to the next and washes off excess dye or mordant.

5. Dehydration and clearing

   Dehydration is done in graded alcohols or acetones progressing from 70% to absolute alcohol or acetone. In case the water has not been completely removed the section gives a milky appearance after the first dip in xylol. Such sections should be returned to absolute alcohol and the process repeated.

6. Cover slipping and mounting

   Clean the section carefully with a clean blotting paper and then place a drop of mountant on the cover slip. Invert the slide over the cover slip and lower it so that it just makes contact with the cover slip. Turn the slide over and then lay it on a flat surface to allow the mountant to spread. Ensure that there are no air bubbles and no excessive pressing (to prevent damage to the section).

Specific histopathological stains are chosen for specific tissues based on the principle that different dyes stain different cellular components. The routine histopathological stain is that of hematoxylin and eosin (H&E). Other stains are referred to as "special stains" because they are employed in specific situations according to the diagnostic need to demonstrate the specific components of tissue e.g. collagen, elastic tissue, reticulin, pigments, nuclear material etc.

Haematoxyl in is extracted by boiling the wood of the logwood tree (Hematoxylin campechianum), and sold commercially as a crude mixture of hematoxyl in and other, unidentified substances. It is available as a tan colored powder which has poor solubility in water and somewhat bettersolubility in ethyl alcohol. The active substance is not hematoxylin, but its oxidized product, hematein, which can be obtained by oxidizing the crude hematoxylin. Oxidation (also called ripening) can be achieved naturally by leaving the hematoxylin exposed to light and air for six weeks to several months. Artificial ripening or oxidation is faster and achieved by adding a wide variety of oxidizing agents, the two most common being sodium iodate and mercuric oxide. The addition of glycerin to several formulae is said to guard against over-oxidation and perhaps to retard fungal growth.

Classification of haematoxylins according to the mordant used:

I. Alum haematoxylins:

The three main alum haematoxylin solutions employed in histopathology are Ehrlich's haematoxylin, Harris's haematoxylin, and Mayer's haematoxylin. The preferred name for these hematoxylins is haemalum as the actual active dye-metal complex contains hematein, a product of oxidation of heamatoxylin and aluminium ions. Alum hematoxylin solutions impart to the nuclei of cells a light transparent red stain that rapidly turns blue on exposure to any neutral or alkaline liquid.

Three main items are needed to produce an effective nuclear staining alum haematoxylin. These are:-

- Haematoxylin or hematein, as the dye.
- An aluminum salt, as the mordant.
- A solvent, usually water.

In addition to these three items, other ingredients may be added. These are not essential but modify the behavior in some fashion. They include: –

- An oxidizing agents to convert haematoxylin (the dye precursor), to hematein, (the dye).
- Acids to balance out the pH, aimed at extending the shelflife of the solution.

- Stabilizers, which inhibit further oxidation once the hematein has been formed. Extended oxidation can shorten the life of the stain and reduce quality of nuclear staining. Other additions to the solvent include agents which inhibit evaporation or precipitation of the stain.

Most of the routinely used haematoxylins belong to this group. The mordant is an alum in the form of potassium or ammonium alum. They stain nuclei a red color that will change to black when washed with weak alkali. Alkali used may be saturated 1% lithium carbonate or 0.05% ammonia or Scott's tap water substitute (sodium or potassium carbonate 2 to 3 gm, magnesium sulphate 20 gm, distilled water 1000 ml) is used (bluing). These can be used regressively if the section is overstained and then differentiated in acid alcohol followed by bluing or progressively when the section is stained for a predetermined time to stain nuclei adequately but leave the back ground tissue relatively unstained. Types include:

- Ehrlich's hematoxylin

Hematoxylin 2 g

Absolute alcohol 100 ml

Distilled water 100 ml

Glycerol 100 ml

Glacial acetic acid 10 ml

Ammonium or potassium aluminium sulphate to saturation – 10 to15 g

Sodium iodate 0.9 g

Dissolve the hematoxylin fully in the alcohol before adding the other ingredients in the order given above. Other additives include "glycerin" which is added to slow the oxidation process and prolong the shelf life of the heamatoxylin and "sodium iodate" which artificially ripens the hematoxylin so that it may be used immediately. Alternatively, the stain can be naturally ripened by exposure to warmth and sunlight for approximately two months. The naturally ripened form has a longer shelf life.

- Mayer's hematoxylin

Hematoxylin 1 g

Distilled water 1000 Ml

Potassium or ammonium alum 50 g

Citric acid 1g

Chloral hydrate 50 g

Sodium iodate 0.2 g

Hematoxylin is dissolved in distilled water using gentle heat. Potassium or sodium alum and sodium iodate are then added. This is followed by chloral hydrate and citric acid while continuing heating. The stain is ready for immediate use when cool.

- Harris hematoxylin

Hematoxylin 2.5 g

Absolute alcohol 50 ml

Distilled water 500 ml

Ammonium or potassium alum 50 g

Mercuric oxide (yellow) 1.5 g

Dissolve the hematoxylin in absolute alcohol. Dissolve the alum in warm water. Mix the two solutions, rapidly bring to the boil and slowly and carefully add the mercuric oxide. The stain is then rapidly cooled by immersing the flask into iced water. 20 ml of glacial acetic acid may be added when the solution is cold to ensure a sharper nuclear staining; this must be added just before use and the stain filtered.

Gill's hematoxylin

Distilled water 730 ml

Ethylene glycol 250 ml

Hematoxylin 2 gm

Sodium iodate 0.2 gm

Aluminium sulphate 17.6 g

Glacial acetic acid 20 ml

Combine the reagents in the order given above and mix with a stirrer for 1 hour at room temperature. The stain can be used immediately.

- Carazzi's hematoxylin

Hematoxylin 0.5 g

Potassium iodate 0.01 g

Potassium alum 25 g

Glycerol 100 ml

Distilled water 400 ml

Combine hematoxylin and glycerol. Dissolve the potassium iodate in a little of the water and prepare the alum using the remainder. Mix the haematoxylin and alum solutions and then carefully add the potassium iodate.

- Delafield's hematoxylin

Hematoxylin stock solution: 1.0 g haematoxylin dissolved in 50 ml absolute ethanol

Sodium iodate stock: 10 g sodium iodate dissolved in 100 ml distilled water

Delafield stock solution:

50 ml hematoxylin stock plus 2 ml sodium iodate stock, mix and wait for 10 min, add 160 ml ammonium alum stock, mix vigorously for 1 min, add 40 ml glycerol. This has similar longevity to Ehrlich's haematoxylin

- Cole's hematoxylin

This is a chemically ripened alcoholic iodine solution.

Hematoxylin stains have a variety of formulations which can be used for different purposes. Usage depends on whether progressive or regressive staining is being used. Also, in situations where haematoxylin staining is followed by acidic stains, Iron haematoxylin is preferred as it resists decolourization by these counter stains. The formulations differ mainly with regard to the mordant and the oxidizer used.

Following are the various formulations commonly used:

1. Harris's haematoxylin – Excellent nuclear stain which is used as a regressive stain in histopathology and progressive stain in exfoliative cytology; it is the preferred haematoxylin in cytology.
2. Mayer's haematoxylin - Most commonly used haematoxylin (staining is progressive).
3. Iron haematoxylin - To be used in situations where counter stains have a strongly acidic character.
4. Phosphotungstic acid haematoxylin (PTAH) - This is exclusively used to demonstrate neurological fibers, muscle striations and fibrin.
5. Others - Ehrlich's and Delafield haematoxylins. These may be used in place of Mayer's haematoxylin ripening is achieved by exposure to light for 6 to 8 weeks.

II. Iron haematoxylins

- Used both as oxidizing agents and mordant.
- Contain ferric chloride and ferric ammonium sulfate.
- Over oxidation of haematoxylin is a problem so separately prepare and reconstitute minutes before use or use them consecutively.
- They are capable of demonstrating detailed tissue structures but techniques are time consuming.
- Usually require a differentiation stage which requires microscopic control.

Include:

1. Weigert's haematoxylin: Ferric chloride is used as a mordant. This is mostly used when acidic staining solutions are to be applied (e.g., Van Gieson's staining). Staining time is 15-30 minutes. It is indicated for neural tissue.

   Weigert's Iron Hematoxylin Solution:

   Stock Solution A:

   Hematoxylin ---------- 1 g

   95% Alcohol ---------- 100 ml

   This is allowed to naturally ripen for 4weeks before use.

   Stock Solution B:

   30% Ferric chloride in water---------- 4 ml

   Distilled water----------95 ml

   Hydrochloric acid, concentrated----- 1ml

   Weigert's Iron Hematoxylin Working Solution:

   Mix equal parts of stock solution A and B. This working solution is stable for 3-6 months.

2. Heidenhain's haematoxylin: This is the most intensely staining heamatoxylin which ues ferric ammonium sulphate as an oxidant/mordant and differentiating fluid. It is used to demonstrate mitochondria, muscle striations, myelin and chromatin, all of which stain grey-black.

   Heidenhain's Hematoxylin

   Hematoxylin solution:

   Hematoxylin ---------- -- 0.5 g

95% Alcohol ---------- 10 ml

Distilled water ---------- 90 ml

Iron solution:

Ferric ammonium sulfate	5g
Distilled water	90 ml

3. Verhoeff's haematoxylin: Verhoeff's technique uses a solution of iron haematoxylin to which iodine has been added. Initially developed as a stain for elastic fibers, it can also be used in electron microscopy and to demonstrate nuclei and myelin. Elastic fibers and nuclei stain black to blue-black, cytoplasm and muscle stain yellow and collagen stains red.

   The staining technique is regressive (ferric chloride is used as a differentiator) and unless carried out carefully, stain can be removed from the smaller fibers. The function of iodine is unclear. It is thought to convert haematoxylin to hematein and also prolong the staining life of the solution.

4. Loyez haematoxylin: Ferric ammonium sulfate is used as a mordant. It is used for demonstration of myelin.

III. Tungsten haematoxylins

There are many preparations of tungsten haematoxylins (tungsten mordanted with haematoxylin), of which, the most widely used one is that developed by Mallory (Mallory's Phosphotungstic Acid Haematoxylin). Although, originally formulated for ne uroglial fibers, this PTAH also stains nuclei, centrioles, mitochondria, fibrin, red blood cells, cardiac and skeletal muscle striations, myelin and some microorganisms, all of which stain blue, while collagen, reticular and elastic fibers, cartilage and bone matrix appear reddish-brown. The best method of preparation is light and air induced oxidation. This method of ripening takes months but remains stable for a long time.

Other mordants include other metallic salts which form colored complexes with haematoxylin like chromium, molybdenum, copper and lead.

Eosin is an acidic xanthine dye with an affinity for cytoplasmic components of the cell. There are a variety of eosins that can be synthesized for use. Eosin is easier to handle than haematoxylin. The only problem one encounters is over staining, especially with decalcified tissues. Some of the more common ones are:

- Eosin
- Eosin Y (most widely used and is soluble in both alcohol and water)

- Alcoholic eosin Y
- Eosin B
- Eosin- phloxine
- Picro-eosin

Common Terminology pertaining to staining:

Basophilic substances: Substances which stain with basic dyes

Acidophilic substances: Substances which stain with acid dyes

Vital staining: Staining of living or viable cells, either in the body (in vivo) or in a laboratory preparation (in vitro).

Progressive staining: Stain is applied to the tissue just long enough to reach the proper end point. To achieve this the slides are examined at regular intervals to determine when staining is appropriate (dark enough but not too dark). Washing or decolorization is not required because there is no over staining of tissue constituents.

Regressive staining: Tissue is first overstained and then the excess stain is removed from all but the structures to be demonstrated. This process is called differentiation and should always be controlled under microscope.

Progressive haematoxylins are generally less concentrated and slow working to prevent overshooting of the endpoint. Regressive haematoxylins are more concentrated and can achieve over staining in a matter of less than a minute, while differentiation (removal of excess stain) can be done in a few seconds. Regressive procedures are therefore faster and more convenient than the progressive ones.

Decolorization: Decolorization is partial or complete removal of stain from tissue sections. When the colour is removed selectively, from all tissue except the target tissue, under microscopic control, it is called differentiation.

Mordants: Substances that facilitate certain staining reactions to take place by forming links between the tissue and the stain are labeled mordants. The link is referred to as a “lake”. The dye is not capable of binding to and staining the tissue in the absence of these links. Examples include ammonium and potassium alum for haematoxylin.

Bluing: Most haematoxylin solutions, which stain regressively, require differentiation, i.e. treatment with an acid reagent to remove excess stain from tissue section. The tissue section is subsequently treated with an alkaline solution to neutralize the acid and restore a blue colour to the tissue. This process is called bluing and it can be achieved by treating tissue

section with either ammonia vapor, 2. 5% ammonium hydroxide for 2 minutes, running tap water for 10 minutes or Scott's tap water substitute (TWS) for 2 minutes.

Metachromatic staining: There are certain basic dyes belonging to aniline group that will differentiate particular tissue components by staining them a different color to that of original dye. This phenomenon is known as metachromasia.

H&E staining

Procedure: Nuclei are first stained blue with a hematoxylin solution and then counterstained with a xanthene dye, e.g., eosin Y, eosin B or erythrosine B. In the latter process the cytoplasm, collagen, keratin and erythrocytes stain red.

Steps:

1. Deparaffinize sections.
2. Hydrate in graded alcohols (absolute alcohol, followed by 95% and 70% alcohol).
3. Wash briefly in distilled water.
4. Stain in Harris haematoxylin solution for 2-5 minutes.
5. Rinse in running tap water.
6. Differentiate in 1% HCL in 70% alcohol for 30 seconds.
7. Rinse in running tap water.
8. Blue in 0.2% ammonia water or saturated lithium carbonate solution for 30 seconds to one minute.
9. Counterstain in eosin-phloxine B solution (or eosin Y solution) for 30 seconds to 1 minute.
10. Dehydrate through 95% alcohol, and absolute alcohol.
11. Clear in two changes of xylene, 5 minutes each.
12. Mount with xylene based mounting medium.

Results:

- Nuclei --------Blue-black
- Cytoplasm --------Varying shades of pink
- Muscle fibers------ Deep pink to red
- Fibrin --------- Deep pink
- Red blood cells ----------Orange/red

Special stains in histopathology:

Special stains are different from the routine stains in the sense that they are "targeted" towards identification of a single cell or tissue constituent. They help in morphological characterization of the tissue as they give different colors to different specific elements of the specimen. This differential staining is due to the different physical, chemical and solubility properties of cell constituents. While putting up a special stain it is always advisable technically to put up a control on tissue which is known to be positive for the target for staining.

Stains for microorganisms

On H&E stain bacteria appear as blue rods or cocci and their colonies appear as fuzzy blue areas. Gram stain is used to further characterize them just like in microbiology except that neutral red is used instead of Safranin. Gram positive organisms usually stain well, but gram negatives do not. Fungi stain blue with H&E and pinkish red with PAS. Methenamine silver stain is the more appropriate stain for fungi. A Giemsa stain can demonstrate organisms causing Leishmania (LD) bodies in tissue sections. Spirochetes can be demonstrated by the Warthin-Starry stain.

a) AFB (acid fast bacilli) stain:

Principle: Carbol-fuchsin stains the lipid walls of acid fast organisms such as M. tuberculosis. The most commonly used method is the Ziehl-Neelsen (ZN) method, though there is also a Kinyoun's method and a fluorescent method Mycobacterial cell walls contain a waxy substance composed of mycolic acids (ß-hydroxy carboxylic acids with chain lengths of up to 90 carbon atoms). This lipid capsule of the acid-fast organism takes up Carbol-fuchsin and resists decolourization with a dilute acid rinse. The degree of acid fastness is directly related to the carbon chain length of the mycolic acid found in any particular species of mycobacteria.

Any routine fixative can be used with the exception of Carnoy's fluid which tends to remove lipid from the bacteria making them non-acid-fast.

Reagent Formulae

1. Carbol fuchsin
   Basic fuchsin.....1 g
   Absolute ethyl alcohol ..... 10 ml
2. Acid alcohol (3%hydrochloric acid in 70% ethyl alcohol).
3. Methylene blue solution

Methylene blue 1g
Distilled water..... 80 ml
Glacial acetic acid. ..... 1 ml
Absolute alcohol… ..... 20 ml

Method

1. Pre-heat a coplin jar containing the working solution in a 58 -60ºC water bath for 10 minutes.
2. Deparaffinize the sections and bring to water.
3. Stain in the working solution in the water bath for 15 minutes.
4. Transfer the coplin jar containing slides in running cold tap water for 2 minutes.
5. Wash the slides in running water for 1 minute.
6. Differentiate in acid alcohol until the tissue becomes very pale pink in color.
7. Wash briefly in water to remove the acid alcohol
8. Counterstain with methylene blue for 15 to 30 seconds.
9. Wash in water, dehydrate, clear and mount in DPX.

Results

- Acid fast bacilli..... Red
- Nuclei. .....Blue
- Other tissue constituents. ..... Blue

Modified Ziehl-Neelsen method for Mycobacterium leprae:

Leprosy bacillus is not as acid fast as Mycobacterium tuberculosis, therefore 1% acid alcohol is used for differentiation.

b) Fite-Faraco Stain for Leprosy Bacillus:

As already stated, the leprosy bacillus is much less acid and alcohol fast than Mycobacterium tuberculosis. therefore in this method 10% sulphuric acid is used as a decolourizer in place of the acid / alcohol solution. Also, the sections are deparaffinized using peanut oil/xylene mixture, which helps to protect the more delicate waxy coat of the organisms.

Reagent Formulae

1. Xylene/peanut oil
   Xylene ---------- 2 parts

Peanut oil ---------- 1 part

2. Carbol fuchsin
3. Sulphuric acid 10%

   Distilled water---------- -- 90.0 ml

   Conc. sulphuric acid ------ -- 10.0 ml
4. Acetified methylene blue

Method

1. Deparaffinize sections in a mixture of two parts xylene/one part peanut/vegetable oil for 15 minutes.
2. Blot dry and wash in water.
3. Stain in Carbol fuchsin solution for 20 minutes without heating.
4. Wash in running tap water.
5. Differentiate in 10% sulphuric acid for 2 minutes (or till pale pink).
6. Wash in running tap water, rinse in distilled water.
7. Counterstain in methylene blue for 20 seconds.
8. Wash and blot dry.
9. Clear in xylene.
10. Mount in DPX.

Results

- Leprosy bacilli ..... magenta
- Nuclei, background ..... blue
- Red blood cells... .....pale pink

c) Gram stain

Principle: It is based on the chemical and physical properties of the cell walls of microorganisms. It basically detects peptidoglycan, which forms about 50% to 90% of the cell envelope of Gram positive bacteria. Gram-negative bacteria have a thin layer of peptidoglycan (forming 10% of cell envelope), which is stained pink by the counter-stain. A Gram positive stain results in a purple/blue color while a Gram negative stain results in a pink/red color. Crystal violet (CV) dissociates in aqueous solutions into CV^+ and chloride (Cl^-) ions. The CV^+ ion react with the negatively charged components of bacterial cells and stains the cells purple. Iodine reacts with CV^+ to form crystal violet and iodine complexes (CV–I) within the inner and outer layers

of the cell. In the presence of a decolorizer like alcohol or acetone, a Gram-negative cell loses its outer lipopolysaccharide membrane, and the inner peptidoglycan layer is left exposed. The CV–I complexes are washed from the Gram-negative cell along with the outer membrane. On the other hand, the large CV–I complexes become trapped within the Gram-positive cell due to the multilayered nature of its peptidoglycan. Thus, after decolorization, the Gram-positive cell retains its purple color and the Gram-negative cell loses its purple color. Safranin or basic fuchsin, which are positively charged dyes, are then used as counterstains to give decolorized Gram-negative bacteria a pink or red color.

Reagent Formulae

1. Crystal violet stain (Reagents may be available as commercial solutions or prepared).
   - Crystal violet 2.0 g
   - 20 ml of 95% alcohol
   - Ammonium oxalate 0.8 g
   - Distilled water 80 ml

   The solution is prepared by dissolving the dye in alcohol, the ammonium oxalate in the distilled water, and mixing the two solutions together. The mixture is stable for two to three years.

2. Gram's iodine
   - Iodine crystals 1 g
   - Potassium iodide 2 g
   - Distilled water 300 ml

   Dissolve the potassium iodide in 2 to 3 ml only of the distilled water - the crystals will dissolve and the solution will become very cold. Dissolve the iodine crystals in the concentrated potassium iodide solution. Dilute the mixture with the remainder of the distilled water.

3. Acetone

4. Safranine obtained commercially

Method

1. Bring sections to water
2. Flood the slide with filtered crystal violet for 10 sec
3. Wash briefly in water to remove excess crystal violet

4. Flood the slide with Gram's iodine for 10 sec
5. Wash briefly in water
6. Decolourize with acetone (1-2 seconds)
7. Wash in tap water
8. Counterstain with safranine for 15-30 seconds. Neutral red can also be used as a counterstain.
9. Dehydrate in absolute alcohol, clear and mount.

Results

Gram positive organisms, Fibrin ..... blue/black

Filaments of Nocardia and Actinomyces mycelia..... blue

Gram negative organisms, Actinomyces clubs. .....red

Nuclei.....red

Stains for Connective Tissue

Trichrome Staining

Trichrome staining is a general term for a number of techniques (e.g. Masson's trichrome, van Gieson's stain, Phosphotungstic acid heamatoxylin, Gomori's one-step trichrome and Verhoeff method for elastic fibers). The trichrome stains are so named as they use three dyes to impart three different colors to tissue elements, such as collagen, fibrin, muscle, red cells, cytoplasm and nuclei respectively. The three most common trichrome stains are Masson's trichrome, Gomori's one-step trichrome and the van Gieson's stain. Of these, the first two are used in the majority of laboratories.

Trichrome staining is based on the principle that different tissues have different molecular sizes and porosity,in conditions of controlled pH. In the routine H&E staining method collagen, elastic fibers and smooth muscle appear pink or reddish in color. In the van Gieson's stain, collagen, which is more permeable than muscle is exposed to acid aniline dyes (acid fuchsin) and picric acid. Collagen gets colored by the dye with the larger molecules. If applied sequentially, the tissue with lowest permeability will retain the first dye. The small molecules of picric acid penetrate all the tissues rapidly, however, are retained only by red cells and muscle (tissue with lowest permeability) while collagen (tissue with greater permeability) releases acid fuchsin to be replaced by aniline blue, a dye with larger molecules.

The Masson's trichrome stain uses iron hematoxylin (which stains nuclei black); Biebrich scarlet (which stain cytoplasm red) and aniline blue or aniline light green (which stain collagen blue or green), in that sequence respectively.

The Gomori's One-Step method on the other hand uses a single solution which contains all the dyes in a single solution together with phosphotungstic acid and glacial acetic acid. The red color in this Gomori's one-step method is imparted by chromotope 2R.

Trichrome stains are used for

1. Histological diagnosis of disorders associated with abnormalities of collagen.
2. Differential staining of collagen and other connective tissue.
3. Differential collagen and smooth muscle in tumors.

Masson's trichrome staining protocol for collagen fibers :

Tissue is fixed in10% formalin or Bouin's solution as it improves staining quality.

Solutions and Reagents:

1. Bouin's Solution:

   Picric acid (saturated)---------- 75 ml
   Formaldehyde (37-40%) ---------- 25 ml
   Glacial acetic acid ---------- 5 ml
   Mix well.

2. Weigert's Iron Haematoxylin Solution:

   Stock Solution A:

   Haematoxylin----------1 g
   95% Alcohol ---------- 100 ml

   Stock Solution B:

   29% Ferric chloride in water --------- 4 ml
   Distilled water---------- 95 ml
   Hydrochloric acid, concentrated ---- 1ml

Weigert's Iron Haematoxylin Working Solution:

Mix equal parts of stock solution A and B. This working solution is stable for 3 months (no good after 4 months)

Biebrich's Scarlet-Acid Fuchsin Solution:

Biebrich's scarlet, 1% aqueous----------90 ml

Acid fuchsin, 1% aqueous ----------10 ml
Acetic acid, glacial ---------- 1 ml

Phosphomolybdic-Phosphotungstic Acid Solution:

5% Phosphomolybdic acid ---------- 25 ml
5% Phosphotungstic acid ---------- 25 ml

Aniline Blue Solution:

Acetic acid, glacial ---------- 2 ml
Distilled water----------100 ml

1% Acetic Acid Solution:

Acetic acid, glacial ---------- 1 ml
Distilled water----------99 ml

Procedure:

1. Deparaffinize and rehydrate through 100%, 95% and 70% alcohol.
2. Rinse running tap water for 5-10 minutes to remove the yellow color due to Bouin's solution.
3. Stain in Weigert's iron haematoxylin working solution for 10 minutes.
4. Rinse in running warm tap water for 10 minutes for blueing.
5. Wash in distilled water.
6. Stain in Biebrich's scarlet-acid fuchsin solution for 10-15 minutes.
7. Wash in distilled water.
8. Differentiate in phosphomolybdic-phosphotungstic acid solution for 10-15 minutes or until collagen is not red.
9. Transfer sections directly (without rinse) to aniline blue solution and stain for 5-10 minutes. Rinse briefly in distilled water and differentiate in 1% acetic acid solution for 2-5 minutes.
10. Wash in distilled water.
11. Dehydrate very quickly through 95% ethyl alcohol, absolute ethyl alcohol (these step will wipe off Biebrich's scarlet-acid fuchsin staining) and clear in xylene.
12. Mount with resinous mounting medium.

Results:

Collagen ---------- blue

Nuclei ----------black

Muscle, cytoplasm, keratin ----------red

Stains for Carbohydrates

a) PAS (Periodic Acid Schiff) Staining

Principle: PAS staining is commonly used in histology to demonstrate carbohydrates. Periodic acid treatment oxidizes the carbohydrates to form dialdehydes. It cleaves the C-C bonds where carbon atoms have adjacent OH or OH and amino groups (1,2 glycols or their amino or alkylamino derivatives). These resulting dialdehyde structures then react with Schiff's reagent (a leucofuchsin containing three dyes of triamino-triphenyl methane type, namely Rosanilin, Pararosalin and Magenta II) to restore the quinoid chromophoric grouping, giving a magenta colored final product to the PAS positive substances.

Basic fuchsin So2 (reduction) Leucofuchs

Leucofuchsin Aldehyde (reoxidation) Magenta colored compound

Leucofuchsin Magenta colored compound

Leucofuchsin is a colorless compound owing to loss of quinoid structure due to destruct tion of chromophoric double bond).

Fixation: 10% formalin.

Solutions and Reagents:

1. Periodic Acid Solution:

Periodic acid ----------1 g

Distilled water----------200 ml

Other oxidants include chromic acid, potassium permanganate, lead tetraacetate and lead bismuthate. Periodic acid is the oxidant of choice as it is most specific for 1,2 glycols or their amino and alkyl amino derivatives. Also it does not further oxidize the aldehydes so formed to carboxylic acid.

2. Schiff Reagent:

Dissolve 1g basic fuchsin in 200ml of boiling distilled water, just after removal of the flask of water from flame. Allow the solution to cool to 50 degrees C and add 2 g of potassium

metabisulphite. Mix and further cool to room temperature and add 2 ml concentrated hydrochloric acid, mix and add activated charcoal and leave overnight at room temperature. Filter through number one Whatman filter paper to get a clear or pale yellow solution. Store in a dark container at 4 degrees C (Schiff's reagent deteriorates rapidly if not kept in a closed container. When a pinkish discoloration appears, discard the reagent).

3. Harris's or Mayer's Haematoxylin Solution

Procedure:

1. Deparaffinize and hydrate to water. Duplicate sections can be dewaxed and hydrated in case diastase treatment is required.
2. Oxidize in periodic acid solution for 5-10 minutes (on longer oxidation proteins and acid mucopolysaccharides will also become PAS positive).
3. Rinse in distilled water and completely dry.
4. Place in Schiff reagent and cover for 15 minutes (sections become light pink during this step).
5. Wash in running tap water for 5 minutes (immediately sections turn dark pink in color).
6. Counterstain in Harris's haematoxylin for 1 minute. Subject the section to differentiation and blueing as usual.
7. Wash in tap water for 5 minutes.
8. Dehydrate and cover slip using a synthetic mounting medium.

Diastase digestion

1. Bring sections to water.
2. Treat with human saliva or with a freshly prepared solution of 0.1% malt diastase in distilled water for 30 minutes at 37degrees.Test for Schiff reagent: Pour 10 ml of 37% formalin into a watch glass. To this add a few drops of the Schiff reagent to be tested. A good Schiff reagent will rapidly turn a red-purple color. A deteriorating Schiff reagent will give a delayed reaction and the color produced will be a deep blue-purple.

Results

Simple polysaccharides, neutral mucosubstances, some macro mucosubstances and basement membranes are PAS positive.

Glycogen, mucin, reticulin, colloid, amyloid ----------Magenta

Fungi ---------- Magenta

Background ---------- Blue

b) **Mucins:** Mucin is a term synonymously used for "mucopolysaccharides" coined by Meyer in the nineteen thirtees. They contain polysaccharides covalently linked to a varying amount of protein and were subsequently classified into epithelial mucins and connective tissue mucins. Mucins contain free hexose groups and certain acidic moieties which influence their histochemical activity. They show a eosinophilic appearance on H&E staqining except with Ehrlich's heamatoxylin (which gives a bluish color). Mucins include acidic mucins (strongly sulphated, weakly sulphated, carboxylated sialomucins, sulphated sialomucins and carboxylated mucins) and neutral mucins. Earlier techniques to demonstrate them used iodine and metachromatic staining, gradually new techniques evolved,e.g., Carmine by Best, PAS by Mc Mannus, Alcian blue by Steedman and aldehyde fuchsin by Gomori. Mucicarmine stain was originally devised by Mayer and later modified by Southgate who added aluminium hydroxide to improve clarity of the staining.

Mayer's Mucicarmine Method

Tissue should be fixed in 10% Buffered Neutral Formalin

Staining Procedure

1. Deparaffinize and hydrate to water.
2. Stain in working Weigert's Hematoxylin about seven minutes.
3. Wash in running water for 10 minutes.
4. Stain in the working Mucicarmine solution (mix 1 volume of Mucicarmine Stock, plus 4 volumes of tap water) for 60 minutes of longer.
5. Give 2 or 3 changes in distilled water.
6. Counterstain in Metanil Yellow, 0.25%, for 15 seconds to 1 minute or longer.
7. Rinse for 2-4 seconds in distilled water.
8. Dehydrate, clear and mount.

Results

- Mucins. ..... magenta
- Nuclei. ..... blue
- Capsule of Cryptococcus neoformans. ..... magenta

Southegate's Mucicarmine staining

Tissue can be fixed in formol saline.

Principle: Aluminium combines with carmine to give rise to a positively charged carmine-aluminium complex which binds readily with negatively charged acid mucins. Strongly

sulphated mucins are variable in their reaction; neutral mucins do not stain at all, while the other acidic mucins (particularly hyaluronic acid) stain strongly.

Reagent Formulae

Mucicarmine solution

Carmine	1.0 g
50% alcohol	100 ml
Aluminium hydroxide	1.0 g
Anhydrous aluminium chloride	0.5 g

Grind the carmine to a fine powder, place 1g of the powdered carmine and 1g of dry aluminium hydroxide in a 500 ml Erlenmeyer flask. Add 100 ml of 50% alcohol, and mix. Add the aluminium chloride and mix. Bring to the boil, and boil gently for two to three minutes. Cool and filter. Store at 4°C. The solution keeps for six months or so.

Procedure:

1. Bring sections to distilled water
2. Stain nuclei with alum haematoxylin for 2 minutes (Ehrlich's haematoxylin should not be used as it stains the mucins)
3. Wash in running tap water
4. Differentiate in acid alcohol
5. Rinse in tap water
6. Blue in Scott's tap water substitute
7. Wash in running tap water
8. Stain with mucicarmine solution 30 mins
9. Wash in running tap water
10. Dehydrate, clear and mount.

Results

- Mucins. ..... Red
- Nuclei. .....Blue

c) Alcian blue stain for acid mucosubstances

Alcian dyes include Alcian blue 8GX, Alcian yellow and mixtures of these.

Principle: Alcian blue stains acidic mucins and acid mucopolysaccharides (sulfated and carboxylated). Alcian blue is a cationic basic dye containing copper phthalocyanins. It stains by forming electrostatic forces between polyanions bearing sulphate or carboxyl groups and

is used to react with (and thus demonstrate), the ionisable moiety of acidic mucins. A tissue component is more intensely stained if the dye is used at a pH at which the reacting groups are fully ionized. Strongly sulphated mucins stain at a pH less than 1 and weakly sulphated mucins stain optimally at a pH between 2.5 to 1.

Reagent Formulae

1. Alcian Blue pH 2.5

   1% Alcian blue in 3% aq acetic acid

   Alcian blue ---------- ----1.0 g

   Distilled water---------- -- 97.0 ml

   glacial acetic acid ---------- -- 3.0 ml

   Dissolve the dye in the distilled water, add acid, mix well. Filter before use.

2. Alcian blue pH 1.0

   Dissolve 1 gm of alcian blue in 90ml of distilled water and 10 ml of 1N hydrochloric acid

   Hydrochloric acid 1N:

   Distilled water..... 915 ml

   Concentrated hydrochloric acid. ....85 ml

3. Alcian blue pH 0.2:

   Dissolve 1 gm alcian blue in 100mls of 10% sulphuric acid.

   Sulphuric acid 10%:

   Distilled water 90ml

   Concentrated sulphuric acid. 10ml

Method

1. Bring sections to water
2. Stain in the Alcian blue solution for 15 mins
3. Wash well in running tap water for 5 mins. Rinse in distilled water
4. Counterstain with 1% aqueous neutral red stain for one minute
5. Rapidly dehydrate in absolute alcohol, clear and mount.

Results

- Acid mucins…..blue
- Nuclei…..red
- Red cells…..yellow

At pH 2.5 most simple no-sulfated acid mucins..... Blue

At pH 1.0 weakly and strongly sulphated acid mucins. ..... Blue

At pH 0.2 strongly sulphated acid mucins ..... Blue

Alcian Blue/PAS Staining Procedure

Fixation: 10% neutral buffered formalin is the preferred fixative, however, other fixatives like Carnoy's may also be used.

Principle: This method utilizes the properties of both Alcian blue and PAS rections to stain the full range of mucosubstances. Alcian Blue pH 2.5 stains the acid mucin blue while the Schiff's reagent stains the neutral mucins pink to red. Mixtures of the two mucins will appear purple due to the positive reactions with both Alcian Blue and Schiff's reagent.

The the section is first stained with Alcian Blue as the acid mucins which stain do not react when the section is subsequently stained with the PAS method.

d) Grocott's Methenamine Silver Staining Protocol

Fixative: 10% buffered neutral formalin.

Principle: This is useful in demonstrating fungus in tissue sections. Fungal cell wall contains l mucopolysaccharide components which form aldehydes on oxidation with chromic acid. These aldehydes reduce an alkaline hexamine-silver complex to metallic silver, rendering them visible.

Reagent Formulae

1. 2% aqeous. Chromic Acid
   Chromium trioxide (analytical) ---- 10 g
   Distilled water---------- 500 ml
2. Silver solution
   3% methenamine ----------100 ml
   5% silver nitrate ----------5 ml
3. 0.5% aq Sodium chloroaurate (yellow gold chloride)
   Gold Chloride (analytical) ---------- 0.5 g
   Distilled water---------- 100 ml

4. 2% aq Sodium thiosulphate (hypo)
   Sodium thiosulphate ---------- -- 2.0 g
   Distilled water ---------- 100 ml
5. Working light green solution 0.2%
   Light green in ---------- --0.2 gm
   Glacial acetic acid ----------0.2 ml
   Distilled water ---------- 100 ml
   6.5% Borase solution ---------- --7.1% sodium metabisulfite

Procedure:

1. Bring sections to distilled water.
2. Oxidise with 2% aqueous chromic acid at room temperature for 1 hr
3. Wash in running tap water for a few minutes.
4. Treat sections with 1% sodium metabisulphite for 1 min to remove the residual chromic acid
5. Wash in running tap water for 3 mins
6. Rinse thoroughly in distilled water.
7. Place in working silver solution in a water bath at 60°C in the dark for 15 to 20 mins until section turns yellowish-brown
8. Rinse well in distilled water
9. Tone sections with gold chloride 2 mins
10. Rinse in distilled water
11. Treat sections with 2% sodium thiosulphate 2 mins
12. Wash with running tap water 5 mins
13. Counterstain in working light green 15 sec
14. Rinse excess light green off slide with alcohol
15. Dehydrate, clear and mount.

Note: Wear protective clothing, gloves and safety glasses when preparing reagents as silver nitrate and chromic acid are toxic and methenamine is flammable.

Results

- Fungi ----------black
- Background ---------- pale green to yellow

e) Methenamine Silver (Gomori PAMS) Staining Protocol for Reticular Fibers and Basement Membranes

Principle:

The mucopolysaccharide components of the tissue are oxidized to release aldehyde groups. The aldehyde groups then react with the silver nitrate, reducing it to metallic silver, rendering them visible. It is used for renal biopsy interpretation.

Solutions and Reagents:

1. 0.5% Aqueous Periodic Acid
2. Methenamine Silver Stock Solution:

   3% Methenamine ---------- 100 ml

   5% Silver nitrate---------- 5 ml

   Add the silver nitrate solution in small amounts to the methenamine solution, mixing after each addition. Stock solution should be clear for use. Solution is stable for several months if stored at 4ºC in a dark container.
3. Methenamine Silver Working Solution:

   Methenamine silver stock solution -------- 50 ml

   5% sodium borate (borax) solution -------- 5 ml
4. 0.2% Gold Chloride
5. 3% Sodium Thiosulfate
6. Nuclear Fast Red

Procedure:

1. Deparaffinize slides to distilled water.
2. Oxidize in 0.5% periodic acid solution for 15 minutes at room temperature.
3. Wash well in doubly distilled water.
4. Incubate slides in methenamine silver working solution for 30 min to 1 hour at 60 °C.
5. Rinse in distilled water and check microscopically.
6. Rinse in distilled water again.
7. Tone in gold chloride solution for 1 minute.
8. Rinse in distilled water.
9. Treat with sodium thiosulfate solution for 2 minutes.
10. Wash in running tap water for 10 minutes.

11. Counterstain in nuclear fast red or light green for 5 min.
12. Dehydrate, clear and mount.

Results:

Basement membranes, reticular fibers --------black

Nuclear or background ---------- pink or green

Giemsa Staining Protocol for Tissue Sections

Fixation: 10% neutral buffered formalin is the preferred fixative, however, other fixatives like Carnoy's may also be used.

Principle: Giemsa stain is a member of the Romanowski group of stains, which contain a mixture of the basic dye, methylene blue, and the acid dye, eosin. These polychromatic stains provide a wide color range when staining tissues due to their ability to form new substances-metachromatic dyes-particularly on standing in solution or at an alkaline pH. Several variants are known based on thedifference in the degree of oxidation (polychroming) of the methylene blue stain prior to the precipitation.

The Romanowski stains are extremely tedious to prepare, and therefore are mostly purchased as the commercially available stock stains.

Reagent Formulae

1. Giemsa stain, stock solution (commercially obtained)
2. Giemsa stain, working solution

   Giemsa stock solution -------40 drops

   Distilled water---------- 40 ml

   The diluted stain keeps well, but is best made up fresh each time.
3. cetic acid 0.5%

Procedure:

1. Bring sections to distilled water
2. Stain with diluted Giemsa stain
3. Rinse in distilled water
4. Differentiate with 0.5% aqueous acetic acid (for differentiation)
5. Dehydrate, clear and mount

Results

Bile pigments. .....green
Collagen, muscle, bone. .....pale pink
Micro-organisms, fungi, parasites. ..... purplish-blue
Nuclei. .....dark blue to violet
Erythrocytes. ..... salmon pink
Cytoplasm… ..... light blue

Stains for Fat

a) Oil Red O staining

Principle: Fat staining is based on the physical property of differential staining (adsorption) of certain dyes like oil red O, Sudan black B, Sudan III, and Sudan IV. Oil Red O is a fat-soluble diazo dye used for staining of neutral triglycerides and lipids on frozen sections.

Fixation: 10% formalin, Formal saline or Formal calcium.

Solutions and Reagents:

1. 0.5% Oil Red O Solution:

   Oil Red O ---------- 0.5 g
   Propylene glycol, 100%----------100 ml

   A small amount of propylene glycol is added to the oil red O and mixed well. The remainder of the propylene glycol is added slowly with continuous stirring. The mixture is heatedup to a temperature of 95 -100 °C. Overheating will result in high background staining. The solution is filtered while still warm and can be stored at room temperature for many years. If precipitate forms in the solution, re-filter.

2. 85% Propylene Glycol Solution:

   Propylene glycol, 100%----------85 ml
   Distilled water---------- 15 ml

3. Gill's or Mayer's Haematoxylin Solution

Procedure:

1. Frozen sections are cut at 5-10 µm thickness.
2. Slides are air dried for 30-60 minutes at room temperature and then fixed in ice cold 10% formalin for 5-10 minutes. They are then rinsed in 3 changes of distilled water and air dried for a few minutes and air dried for a few minutes.

3. Place in absolute propylene glycol for 2-5 minutes to avoid carrying water into Oil Red O.
4. Stain in pre-warmed Oil Red O solution for 8-10 minutes in 60ºC oven.
5. Differentiate in 85% propylene glycol solution for 2-5 minutes.
6. Rinse in 2 changes of distilled water.
7. Stain in Gill's or Mayer's haematoxylin for 30 seconds.
8. Wash thoroughly in running tap water for 3 minutes.
9. Place slides in distilled water.
10. Dehydrate, clear and mount.

Results:

Lipids ---------- red

Nuclei ---------- pale blue

b) Sudan Black staining

Principle: When the Sudan Black stain is dissolved in organic solvent, it exhibits a greater solubility in lipids of frozen tissue than that in original solvents. So during staining, dyes will migrate into lipids from organic solvents resulting in lipid staining.

Fixation: Cryostat sections

Method:

Rinse sections in 70% ethanol.

1. Stain for up to 2 hrs in saturated Sudan Black B in 70% alcohol.
2. Rinse in 70% ethanol and remove excess dye and wash in tap water.
3. Counterstain with neutral red for 2-5 minutes.
4. Wash and mount in glycerin jelly.

Results:

Lipids ---------- Blue/black

Stains for Pigments

a) Perl's Prussian blue staining

Fixation: Neutral buffered formalin is preferred, however, other fixatives may be used, but acidic fixatives, dichromate fixatives, and acidic decalcification fluids are best avoided as they cause progressive loss of ferric ions from tissues.

Principle: Dilute mineral acid hydrolysis releases ferric ions from protein bound tissue deposits of hemosiderin (hemosiderin is ferric hydroxide bound to protein ferritin). These in the presence of ferrocyanide ions, are precipitated as the highly coloured and highly water-insoluble complex, potassium ferric ferrocyanide or Prussian blue. Ferrous ions do not produce a coloured reaction product, and thus are excluded from visualisation.

Reagent Formulae

1. 2% Aqueous hydrochloric acid (Analytical Reagent grade) 25cm3
2. 2% Aqueous potassium ferrocyanide (Analytical Reagent grade to be handled carefully as it is toxic) 25cm3 Mix the above to prepare incubating solution fresh before use.
3. Neutral red stain

    Neutral red ..... 1.0 g

    Distilled water .....100.0 ml

    Glacial acetic acid. .....1.0 ml

Dissolve the dye in the distilled water. Add the acid. Mix well. Filter into the reagent bottle.

Method

1. Bring sections to distilled water.
2. Transfer the sections to the incubating solution for 10 min.
3. Wash well in distilled water, several changes 5 min.
4. Counterstain with filtered 1% aqueous neutral red stain for 1 min (0.1% nuclear fast red in 5% ammonium sulphate for 2-5 min or 0.5% aqueous eosin for 20-60 seconds may also be used).
5. Rinse in distilled water.
6. Rapidly dehydrate in absolute alcohol, clear and mount.

Results

- Ferric salts. ..... deep blue
- Nuclei. .....red
- Erythrocytes. ..... yellow

b) Masson Fontana Staining Protocol for pigments and Argentaffin granules

Principle: Positive argentaffin reactions mean the cells take-up silver and then reduce it to a visible metallic state, without the aid of a reducing agent. Melanin is a brown-black pigment which is normally present in the hair, skin, retina, iris, and certain parts of CNS. This method

is used to demonstrate melanin and other substances which have similar reducing properties, such as argentaffin cell granules and lipofuscin. Argentaffin granules are found in carcinoid tumors and lipofuscin is a wear & tear pigment.

Solutions and Reagents:

1. 10% Aqueous Silver Nitrate Solution:

   Silver nitrate ---------- 10 g

   Distilled water---------- 100 ml

2. Fontana Silver Nitrate Stock Solution

   To 25 ml of 10% Aq. Silver nitrate add ammonium hydroxide drop by drop, until solution precipitates and clears again. Add 10% silver nitrate drop by drop until the solution becomes slightly cloudy or opalescent. Leave solution overnight. Store it in dark at room temperature. Stock solution is stable for approximately 6 months. (CAUTION: CORROSIVE - avoid contact and inhalation).

Fontana Silver Nitrate Working Solution:

Fontana silver nitrate stock solution---- 25 ml

Distilled water---------- 75 ml

Filter is not necessary. Discard after use.

3. Gold Chloride Working Solution:

   Gold chloride solution, 1% stock --------- 10 ml

   Distilled water---------- 40 ml

4. Neutral Red Stain – acidified

Neutral red	1 g
Distilled water	100 ml
Glacial acetic acid	1 ml

Dissolve the dye in the distilled water. Add the acid. Mix well. Filter into the reagent bottle.

Fixation: 10% formalin

Procedure:

1. Bring sections to water.
2. Place slides in Fontana silver nitrate working solution and leave in a 56^{o}C oven for 2 hours. Slides may be checked after 1 hour.
3. Rinse in 3 changes of distilled water.

4. Tone in gold chloride working solution for 1 minute.
5. Rinse in distilled water.
6. Place in 5% sodium thiosulfate solution for 1 minute.
7. Rinse in distilled water.
8. Counterstain with nuclear neutral/fast red solution for 2-5 minute
9. Rinse thoroughly in distilled water twice.
10. Dehydrate, clear and mount.

Technical Points

1. Ideally a known positive control (a nevus for melanin, or small intestine for argentaffin granules) must be used to ensure that the staining protocol is correct.
2. The slide must be repeatedly checked microscopically after every 15 mins in step 2.
3. Slides should be washed well with distilled water. If the neutral red stain is applied directly from tap water, red background staining may be seen.
4. The ammoniacal silver solutions should be neutralized immediately after use with saturated sodium chloride and discarded as they can be inherently explosive.

Results

- Melanin. .....black
- Argentaffin cell granules. ..... black
- Lipofuscin. .....black
- Nuclei. ..... red

c) Von Kossa's Staining Protocol for Calcium

Principle: This is an indirect technique is for demonstrating deposits of calcium or calcium salts. The stain does not directly stain calcium ions; instead, tissue sections are treated with a silver nitrate solution to replace the calcium by silver deposits, which are subsequently reduced by the strong light, and visualized as metallic silver.

Fixation: Formalin fixed, paraffin embedded tissue sections or alcohol fixed, frozen sections.

Solutions and Reagents:

1% Aqueous Silver Nitrate Solution:

Silver nitrate---------- 1 g

Distilled water---------- 100 ml

5% Sodium Thiosulfate:

Sodium thiosulfate ---------- 5 g

Distilled water ---------- 100 ml

0.1% Nuclear Fast Red Solution:

Nuclear fast red ---------- 0.1 g

Aluminum sulfate ---------- 5 g

Distilled water ---------- 100 ml

Aluminum sulfate is dissolved in water. Nuclear fast red is then added and the solution slowly heated to boil and cool. The stain is filtered and a grain of thymol is added to ensure preservation..

Procedure:

1. Deparaffinize and bring sections to water.
2. Rinse in multiple changes of distilled water.
3. Incubate sections with 1% silver nitrate solution in a glass coplin jar placed in front of a 60-100 watt light bulb for 1 hour.
4. Rinse in several changes of distilled water.
5. Remove excess silver with 5% sodium thiosulfate for 5 minutes.
6. Rinse in distilled water.
7. Counterstain with fast red for 5 minutes.
8. Rinse in distilled water.
9. Dehydrate through graded alcohols, clear and mount.

Results:

Calcium salts ---------- black or brown-black

Nuclei ---------- red

Cytoplasm ---------- pink

d) Alizarin Red S Staining Protocol for Calcium

Principle: Alizarin Red S is an organic anthraquinone derivative, which is used to directly identify calcium in tissue sections. Though the reaction is not strictly specific for calcium and magnesium, manganese, barium, strontium, and iron may also be stained, but these elements usually do not interfere with the staining as they are present in insignificant quantities. Calcium is chelated to form an Alizarin Red S-calcium complex.

Fixation: Neutral buffered formalin or alcoholic formalin are used. Solution and Reagents:

Alizarin Red Solution:

Alizarin Red S----------2 g

Distilled water----------100 ml

It is important that the solution is mixed well and the pH is maintained between 4.1- 4.3 with 10% ammonium hydroxide.

Acetone (100%)

Acetone-Xylene:

Acetone (100%)----------50 ml

Xylene ---------- 50 ml

Procedure:

1. Deparaffinize sections and bring them to distilled water.
2. Stain slides with the Alizarin Red Solution for 30 seconds to 5 minutes, and observe the reaction microscopically for development of a red-orange color.
3. Excess dye is removed by blotting sections.
4. Dehydrate, clear in acetone as well as the acetone-xylene mixture and mount.

Results:

Calcium deposits (except oxalate)--------- orange-red

e) Congo Red Staining Protocol for Amyloid

Principle: This technique can be used for the detection of amyloid in formalin-fixed, paraffin-embedded tissue sectionsas well as frozen sections. The amyloid deposits will be stained red and the nuclei will be stained blue.

Fixation: 10% formalin.

Solutions and Reagents:

0.5% Congo red in 50% alcohol:

Congo red -------- 0.5 g

50% Alcohol ---------- 100 ml

1% Sodium Hydroxide:

Sodium hydroxide ----------1 g

Distilled water----------100 ml

Alkaline Alcohol Solution:

1% Sodium hydroxide ---------- 1 ml

50% alcohol ---------- 100 ml

Procedure:

1. Deparaffinize and bring sections to water.
2. Stain in Congo red solution for 15-20 minutes.
3. Rinse in distilled water.
4. Differentiate quickly (5-10 dips) in alkaline alcohol solution.
5. Rinse in tap water for 1 minute.
6. Counterstain with Gill's hematoxylin for 30 seconds.
7. Rinse in tap water for 2 minutes.
8. Dehydrate through graded alcohols.
9. Clear in xylene 2 changes, 3 minutes each.
10. Mount with resinous mounting medium.

Results:

Amyloid, elastic fibers, eosinophil granules -------- red

Nuclei---------- blue

Table 2.2: Commonly used special stains in surgical pathology

S.No.	Stain	Uses
1	AFB stain	Acid-fast bacteria, hair shafts, and actinomyces
2	Alcian Blue	Sulfated (acidic) mucosubstances
3	Congo Red	Amyloid
4	Masson Fontana	Melanin, and argentaffin cells
5	Phosphotungstic acid hematoxylin (PTAH)	Muscle, fibrin and glial tissue
6	Giemsa	Mast cells, bacteria, and parasites
7	GMS	Fungus

8	Gram stain	Blue- gram positive: Bacteria, keratin, Nocardia, Actinomyces mycelia, and fibrin Red- gram negative: Bacteria
9	Perl's Prussian Blue stain	Ferric iron in tissues
10	Mucicarmine	Acid mucopolysaccharides
11	Oil Red O	Fats (red)
12	Sudan black B	Fats (blue black)
13	PAS	Glycogen, mucin, and fungus
14	Reticulin	Reticulin fibers
15	Trichrome	Muscle, and collagen
16	Verhoeff E (VGE)	Elastic fibers
17	Von Kossa	Calcium
18	Warthin Starry	Spirochetes
19	Orcein	Elastic fibers

SPECIAL HISTOPATHOLOGICAL TECHNIQUES

1. Frozen section
2. Electron microscopy
3. Immunohistochemistry
4. Immunofluorescence
5. Autopsy techniques

Frozen section

Intraoperative pathological consultation includes:

1. Frozen section.
2. Cytological preparations (e.g. touch imprints).
3. Aliquoting of the specimen for special studies (e.g. molecular pathology techniques, flow cytometry).

The frozen section procedure is a procedure for rapid analysis of a histopathological specimen for appropriate diagnosis and management of a patient. It is also known as cryosectioning and is most often used in oncological surgery. The quality of the slides produced by frozen section is inferior to the sections obtained after formalin fixed paraffin embedded tissue processing. While diagnosis can be rendered in many cases on the former, the latter is preferred in many conditions for more accurate diagnosis.

The key instrument for cryosectioning is the cryostat, which is an insulated cabinet that houses an instrument to section frozen samples; a rotary microtome and knife (or blade) holder, and a means to freeze samples. Access to the chamber is via a heated sliding window. The normal working chamber temperature is from 0°C to −35°C. Cryosectioning at temperatures lower than −35°C requires the use of a cryogen such as liquid nitrogen. Several types of cryostats are commercially available and can be categorized as follows:

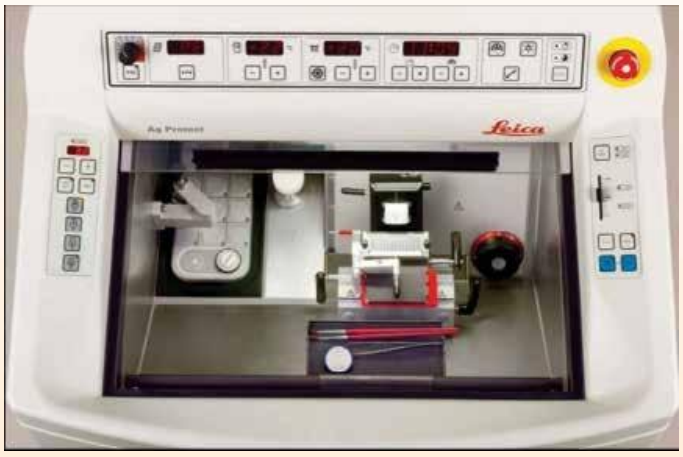


Figure 5.11 : Cryostat - Photo downloaded from "A Practical Guide to Frozen Section Technique" by Stephen R. Peters (Springer)

- Single compressor (chamber cooling only)
- Double compressor (chamber and object cooling)
- Manual sectioning
- Motorized sectioning

The knife (or blade) holder is placed in front of the microtome. Both types of holders (knife and blade) are fitted with antiroll guides, which as the name suggest help to prevent rolling or curling of sections as they are being prepared. The antiroll devices consist of a glass plate supported in a metal frame.

Alternately a cooled brush (an artist's brush) technique can be used to collect and gather sections. In this technique; the brush is used to manipulate the leading edge of the section as it starts to come over the blade or knife edge onto the front surface of the blade. Specimen holders or chucks for cryostats are available in a variety of shapes and sizes.

Figure 5.12: Specimen holders or chucks - Photo downloaded from "A Practical Guide to Frozen Section Technique" by Stephen R. Peters (Springer)

Procedure:

- The selected and trimmed surgical specimen is placed on a metal object disc which is then secured in a chuck and frozen rapidly to about –20 to –30°C. The tissue must be frozen as quickly as possible in order to avoid ice crystal formation resulting in morphological artifacts.
- Due to variation in composition different tissues have different preferred temperatures for processing. Lipid rich substances usually require a lower operating temperature.
- The specimen is embedded in a gel like medium consisting of poly ethylene glycol and polyvinyl alcohol (OCT or "optimal cutting temperature" cryostat sectioning medium) and stored at -80°C until ready for sectioning. This compound is known by many names and when frozen has the same density as frozen tissue. At this temperature, most tissues become rock-hard.
- A small amount of OCT or other suitable frozen section embedding medium (water, bovine albumin and von Apathy's gum syrup) is placed on a cryostat object disk.
- Position the frozen specimen in the center of the object disk and place the disk on the cryobar in the cryostat to begin the quick freeze process.
- Any aerososl refrigerant (e.g., Histo-Freeze) can be sprayed around the periphery of the object disk. As the OCT freezes its appearance changes from a clear gel to white solid substance.
- More OCT is added to cover the specimen top before the disk is frozen solid add and a heat extractor is placed on top of the specimen to (1) rapidly freeze the OCT and tissue and (2) produce a flat embedded surface for easy cutting.
- Place the object disk in the microtome object disk holder and tighten the set screw or clamp.
- Make sure that there is enough clearance between the block and the microtome knife.
- After disengaging the ratchet from the micrometer gear move the block toward the knife edge. Adjust the micrometer setting of the microtome to "trimming" thickness of 15 µm and begin to turn the microtome handwheel; the specimen will advance to make contact with the knife and the surface of the block will be sectioned. This process is termed "trimming" or "facing" the block, and the purpose is to achieve a full face section of the specimen.
- For section cutting adjust the micrometer setting to the desired section value, e.g., 5 µm. Carefully wipe or brush away the surface and edge of the knife and the undersurface of the antiroll plate. Lower the antiroll plate into place and continue sectioning.

- Sections of fresh frozen tissue will adhere to plain glass slides due to the presence of free protein and lipid. Sections of fixed frozen tissue will need to be mounted on coated slides, e.g., poly-l-lysine.
- Subsequently the section is picked up on a glass slide and stained with hematoxylin and eosin.
- The sample preparation time is much more shorter as compared with traditional histology technique.

Electron microscopy

The energy source used in the electron microscope is a beam of electrons. Since the beam has an exceptionally short wavelength, it strikes most objects in its path and increases resolution of the microscope significantly.

Electron microscopes can be of the following types:

- Transmission electron microscope (TEM): It uses electrons passed through the sample to build a picture of the sample's internal structure. It is a more traditional form of electron microscope.
- Scanning electron microscope (SEM): Here an electron beam is projected on the sample. The electrons do not go through the sample but bounce off. This way it is possible to visualize the surface structure of the specimen. The image which is thus formed appears three dimensional.

Transmission electron microscopy, a popular diagnostic adjunct in the 1970s and 1980s, has been largely replaced by immunohistochemical, cytogenetic and molecular techniques. However, EM continues to be used in the diagnosis of:

- Microbial diseases
- Lysosomal storage diseases
- Bullous skin disorders
- Peripheral neuropathies
- Soft-tissue tumors
- Glomerulopathies
- Evaluation of an unknown primary malignancy
- Differentiation of a mesothelioma from an adenocarcinoma

Advantages of EM over optical microscopes:

- Modern electron microscopes can magnify up to two million times. Researchers use it to examine the minutest biological material, cells, molecules, and their physical/crystalline structure, and characteristics of various metals and surfaces.
- It is possible to view the three-dimensional external shape of an object with a scanning electron microscope.

Disadvantages:

- It is extremely expensive and a high-maintenance equipment.
- Requires elaborate sample preparation.
- The prerequisite for a completely dry sample and also the high energy of the electron beam (converting into high radiation) makes it impossible for one to observe living or moving specimens with an electron microscope.
- It is not possible to observe color as electrons do not possess a color.

Immunohistochemistry (IHC)

IHC is the method of localization of antigens in tissue by use of labeled antibodies through antigen–antibody interactions that are visualized by markers such as fluorescent dyes, enzymes, radioactive elements, or colloidal gold.IHC makes it possible to visualize distribution and localization of specific cellular components. Antibody-mediated antigen detection can be done by direct and indirect methods.

Most indirect methods employ inherent binding affinity of avidin to biotin to localize a reporter to target antigen and amplify signal that is detected. IHC target antigens are detected through either chromogenic or fluorescent means.

Chromogenic detection is based on activity of enzymes, most often horseradish peroxidase (HRP) or alkaline phosphatase (AP), which form colored (usually brown); insoluble precipitates upon addition of substrate, such as Diaminobenzidine tetra hydrochloride (DAB) and Aminoethylcarbazole (AEC).

Applications

- IHC can be used for tumor diagnosis in case routine histology is not diagnostic.
- Tumor markers can be used to differentiate between a benign or malignant tumor and identify the cell/site of origin of a metastasis.
- Early or micrometastasis can be difficult to detect using conventional histology but can be diagnosed by IHC.

- It can be used to prognosticate a cancer (localization of oncogenes, tumor suppressor genes, cell proliferation, or cell death/apoptosis markers) or to determine/predict the response of a tumor to therapy (localization of estrogen–progesterone receptors in carcinoma breast).
- IHC tests are now available for diagnosis of infectious diseases, e.g., Herpes virus, Hepatitis B virus (HBV), Cytomegalovirus (CMV), Human Papilloma virus (HPV), and Leptospira.

Immunofluorescence (IF)

- IF involves an antigen–antibody reaction wherein the antibodies are labeled with a fluorescent dye and the antigen–antibody complex is visualized using ultraviolet light (sourced from mercury vapor and xenon gas lamps) in a fluorescent microscope.
- Dyes that absorb ultraviolet rays and emit visible light are called fluorochromes. Fluorochromes such as fluorescein isothiocyanate (FITC) or tetramethyl rhodamine isothiocyanate (TRITC) are chemically conjugated to antibodies that are allowed to bind (directly or indirectly) to the antigen of interest.

  The antigen is thus demonstrated through fluorescence techniques. The emitted fluorescence can then be quantified using a flow cytometer, array scanner, or automated imaging instrument, or visualized using fluorescence or confocal microscopy.
- There are two main methods of IF labeling - direct and indirect.
- The direct immunofluorescence method, whereby the antibody against the molecule of interest is directly conjugated to the fluorescent dye or fluorochrome, is less commonly used.

  In indirect immunofluorescence (sandwich technique), the antibody specific for the molecule of interest (called primary antibody) is unlabeled, and a second anti-immunoglobulin antibody directed toward constant portion of first antibody (called the secondary antibody) is tagged with the fluorescent dye.
- Some substances fluoresce naturally and this fluorescence is called primary fluorescence or autofluorescence, e.g., chlorophyll.

  Those substances that do not fluoresce spontaneously but require induction of fluorochromes are said to emit secondary fluorescence.

Applications:

1. The test is routinely done on renal and skin biopsies for detection of deposits of immunoglobulins, complement and fibrin in glomerulonephritis, and bullous disorders of skin.

2. It can be used for detection of autoantibodies in serum, e.g., antinuclear antibody (ANA) in SLE, anti smooth muscle antibody (SMA) in biliary cirrhosis, antimitochondrial antibody (AMA) in autoimmune hepatitis, autoantibodies to glutamic acid decarboxylase (GAD), the protein tyrosine phosphatase-like molecule, IA-2, and insulin in type 1 diabetes, thyroid autoantibodies in Hashimoto thyroiditis and Grave's disease etc.
3. IF can be applied to analysis of antigens in fresh, frozen, or fixed tissues, and subcellular localization of antigens in tissue culture monolayers.
4. It can be used for detection and localization of the presence or absence of specific DNA sequences on chromosomes patterns of gene expression within cells/tissues.

Autopsy protocol

An autopsy is a medical procedure that consists of a thorough examination performed on a body after death, to evaluate disease or injury that may be present and to determine the cause and manner of death. There are three main types of autopsies

1. Medico-Legal or Forensic autopsies: These are used to determine the cause and manner of death and are generally performed, as prescribed by applicable law, in cases of violent, suspicious or sudden deaths.
2. Clinical or Pathological autopsies: These are performed to diagnose a particular disease or for research purposes.
3. Anatomical or Academic autopsies: These are performed by students of anatomy for study purpose only.

It is preferable that the autopsy examination be performed as soon after death as possible. Therefore, if delay is unavoidable, the body should be placed in the mortuary refrigerator (4°C) or arterial embalming should be carried out without delay provided that toxicological or microbiological examination are not indicated such embalming, well done, is most advantageous and given excellent preservation. Slices of organs taken at autopsy should be thin to facilitate fixation.

Principal Autopsy Techniques:

The four major autopsy techniques differ chiefly in the methods used in removal of the organs and the order in which they are opened.

1. Technique of Virchow: Most widely used method in which the organs are removed one by one. This approach is good when multiple organs are thought to be involved by the disease, for demonstrating pathological changes in individual organs. The disadvantage of this technique is that relationships between various organs may be hard to interpret. Several modifications are available.

2. Technique of Rokitansky: This procedure is characterized by in situ dissection, in part combined with en bloc removal.
3. En mass technique: Thoracic, cervical, abdominal, and pelvic organs are removed en masse and subsequently dissected into organ blocks. This is the best technique for demonstrating the vascular supply of organs and their relationship with one another. The major disadvantage is that it is difficult to handle the organ mass., and performing the autopsy is difficult without an assistant.
4. En Bloc Technique: Thoracic, cervical and abdominal organs, and the urogenital system are removed in functionally related blocks. This procedure is a compromise between the Virchow and en masse techniques, preserving anatomical relationships sufficiently for most cases while being simpler for one person to execute.

STORAGE AND ARCHIVING OF SPECIMENS

Maintenance of histopathology records and surgical and autopsy specimens, is an extremely important aspect of pathology services. This includes filing and storing of paraffin blocks and stained tissue slides usually in order of accession number.

General guidelines for storage and archival in a pathology laboratory

- It is advisable that surgical specimens are stored for a minimum of two weeks to approximately six weeks after a report is issued so that if new clinical information becomes apparent that might require further investigation of the unprocessed tissue, it can be done.
- It is recommended that diagnostic histopathology slides and blocks should be kept for a minimum of ten years. Report duplicates/records/diagrams and copies of any representative images prepared should be kept for at least 20 years.
- All laboratories must retain cytology slides for a minimum of five years.
- Currently, some patients with cancer survive for more than 10 years, and review of previous pathology material may be required for comparison with recurrent tumor or for enrollment in clinical trials. Therefore it is recommended that once the regulated length of time for storage is met, institutions may continue to store pathology specimens based on the room they have for storage.
- The entire archive (paper records, glass slides, and tissue blocks) should be professionally stored in a climate-controlled environment, and its index should be available, preferable in digital format.
- Paper records should be stored as bound volumes. All documents, relevant radiographs, gross specimens and additional material available should be photographed and scanned to be digitally archived as well.

SAFETY IN THE LABORATORY

- Treat all biological materials used in the laboratory as potentially infectious and pathogenic to humans.
- Laboratory coats must be worn by laboratory personnel at all times.
- All open cuts on hands and other exposed skin surfaces must be covered by gloves.
- Long hair should be tied back neatly, away from the shoulders.
- The lab should be well-ventilated and should strictly follow the regulations governing the acceptable limits of formalin and hydrocarbons such as xylene and toluene. If solvents are used during practical sessions, the exhaust fan must be switched on.
- Whenever doing staining procedures ensure that protective gowns, gloves and safety glasses are worn. When 'bringing sections to water' or 'dehydrating, clearing and mounting' always ensure that the exhaust system is turned on.
- The wheel of each microtome must be in the locked position any time the microtome is not being used for cutting sections.
- Utmost care must be taken while handling embedding centers (as they contain paraffin wax at 61oC and above). When using, ensure that you do not burn your fingers.
- A safety data sheet should be maintained for every chemical compound used and it should specify the nature, toxicity, and safety precautions to be taken while handling the compound.
- Proper disposal of hazardous wastes is a must. Tissues that are collected should be stored in formalin and may be disposed by incineration or by putting them through a "tissue grinder" attached to a large sink (similar to a large garbage disposal unit).
- Sharps are to be placed in the sharps containers scalpel blades are to be disposed of in the blade removal system, which are disposed as per recommended guidelines.
- Every instrument used in the laboratory should meet electrical safety specifications and have written instructions regarding its use.
- It is advisable that flammable materials are stored with utmost care in appropriate storage cabinets that are designed for this purpose.
- Fire safety protocols are to be strictly adhered to. Safety equipment including first aid kits, fire extinguishers, fire blankets, and fire alarms should be within easy access. A shower and eyewash should be Specific hazards:

- Bouin's solution contains picric acid, which is only sold in the aqueous state. This is because it is inherently explosive in the dry state.
- Sodium azide, used as a preservative in many kits is also explosive in nature. It should be flushed down the drain with lots of water, or there is a tendency for the azide to form metal azides in the plumbing.
- Benzidine, benzene, anthracene, and napthol containing compounds are carcinogens and should not be routinely used.
- Mercury-containing solutions (Zenker's or B-5) should always be discarded into proper containers. Mercury, if poured down a drain, will form amalgams with the metal that build up and cannot be removed.

Self-Assessment

1. Define histopathology.
2. Enumerate the steps in specimen receiving and accessioning.
3. Enlist the list of equipment required for setting up of a histopathology laboratory.
4. What are the properties of an ideal fixative?
5. Classify fixatives. Which are the most common fixatives used in histopathology. Enumerate their specific uses and disadvantages.
6. Outline the steps in routine processing of histopathology specimens.
7. What is automated processing?
8. Enumerate the steps in specimen mounting for museum preparation.
9. Name the fixation induced pigments. Outline the steps in the methods used to remove them.
10. Enlist the factors affecting the rate of tissue processing.
11. Define dehydration. Enumerate the available dehydrating agents.
12. Define clearing. Enumerate the available clearing agents. Enumerate the criteria for choosing a suitable clearing agent.
13. Define embedding. Which is the most popular embedding medium for histology and why? Enumerate the alternatively available embedding agents.

14. Define microtomy. Enlist the different types of microtomes and knives used in histopathology specimen cutting.
15. Outline the steps in cutting of paraffin embedded tissue.
16. Name the fixatives preferred for processing of bone specimens.
17. Describe the principles of decalcification and enlist the various decalcifying agents used on bone processing. What are their advantages and disadvantages?
18. Enumerate and describe the methods used for checking the end point of decalcification. How does one test for the remaining calcium in decalcifying tissue.
19. Outline the principle of differential staining. Enumerate the steps in routine H&E staining.
20. Classify haematoxylins. Specify the uses of the different types.
21. Define:
    a) Basophilic substances.
    b) Acidophilic substances.
    c) Vital staining.
    d) Progressive staining.
    e) Regressive staining.
    f) Decolorization.
    g) Mordants.
    h) Bluing.
    i) Metachromatic staining.
22. Enumerate the principle of PTAH staining and its applications.
23. Enlist the special stains used for demonstration of microorganisms and describe their principles.
24. Enlist the special stains used for demonstration of connective tissue and describe their principles.
25. Enlist the special stains used for demonstration of carbohydrates and describe their principles.

26. Enumerate the endogenous pigments. Name the special stains used for their demonstration and describe their principles.
27. What are the main indications for a frozen section? Outline the steps in its preparation.
28. What are the principles of electron microscopy? Enlist its types.
29. Enumerate the advantages and disadvantages of electron microscopy over light microscopy.
30. Write briefly on the applications of immunofluorescence techniques in histopathology.
31. Discuss the role of immunohistochemistry in histopathology.
32. What are the different types of autopsies? Write briefly on autopsy protocol.

Central Board of Secondary Education

Shiksha Kendra, 2, Community Centre, Preet Vihar, Delhi-110301
Phone No. : 011-22527183, Fax : 011-22526580
E-mail : voc.cbse@gmail.com
Website : www.cbsevocational.in